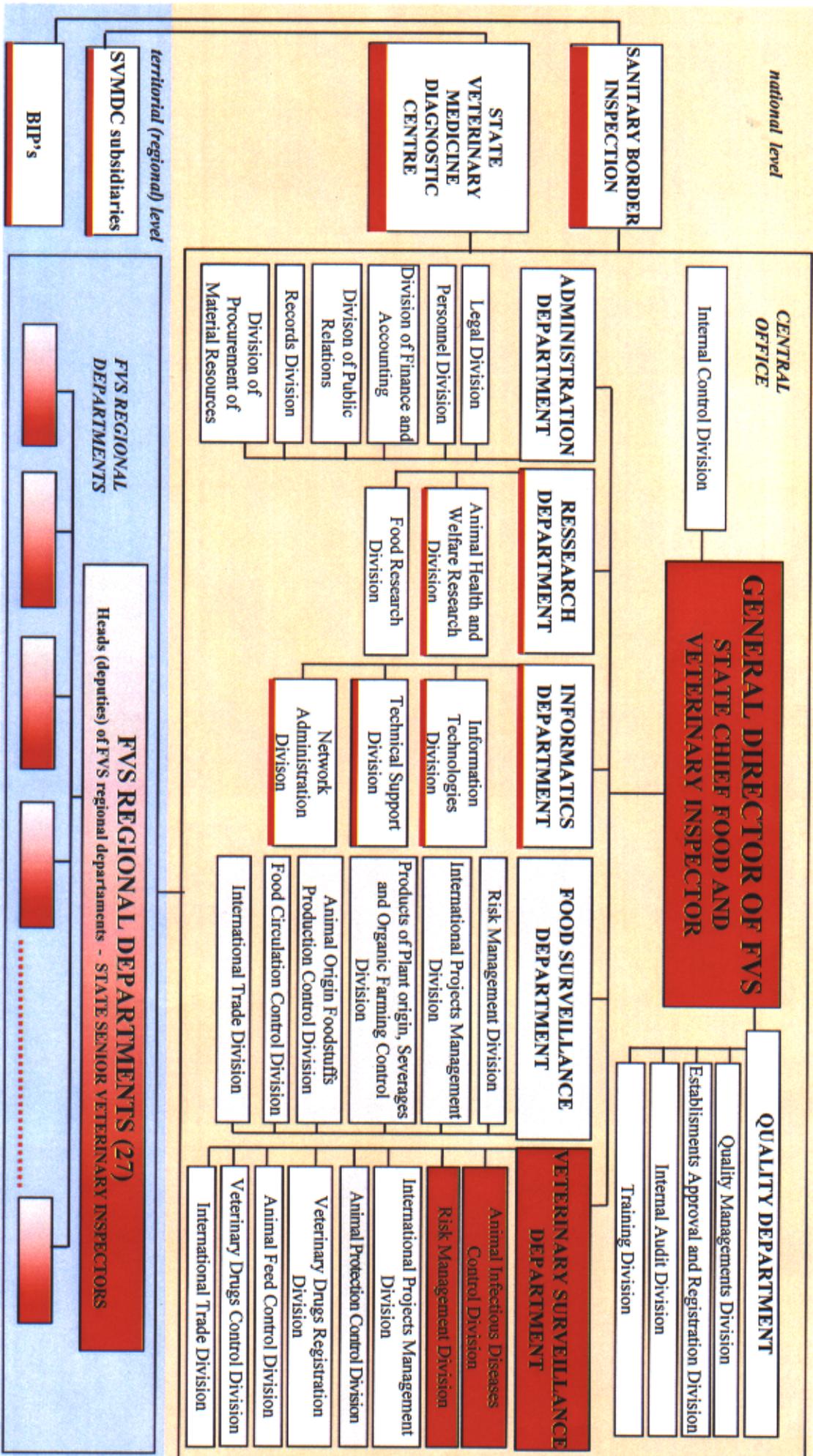


**HEME OF THE FOOD AND VETERINARY SERVICE**

Fig. 1



**DZĪVNIĒKU INFEKCIJAS SLIMĪBU  
VALSTS UZRAUDZĪBAS  
2004. GADA RĪCĪBAS PLĀNS**



**LATVIJAS REPUBLIKAS  
PĀRTIKAS UN VETERINĀRAIS DIENESTS**

APSTIPRINĀTS

Ar Valsts galvenā pārtikas un veterinārā inspektora  
2003. gada 10. decembra rīkojumu Nr. 299.

**DZĪVNIEKU INFEKCIJAS SLIMĪBU VALSTS  
UZRAUDZĪBAS 2004. GADA RĪCĪBAS PLĀNS**

IZDOTS

Pamatojoties uz Veterinārmedicīnas likuma  
16. panta 5. apakšpunktu

**RĪGA, 2003**

**DZĪVNIĒKU INFEKCIJAS SLIMĪBU VALSTS UZRAUDZĪBAS RĪCĪBAS PLĀNS** (turpmāk tekstā – rīcības plāns) ir Pārtikas un veterinārā dienesta izstrādāts un uzturēts dzīvnieku infekcijas slimību valsts uzraudzības pasākumu plāns, ar kura palīdzību praksē realizē dzīvnieku infekcijas slimību kontroli un profilaksi, atbilstoši šo slimību valsts uzraudzības programmām.

**RĪCĪBAS PLĀNS** paredz noteiktu pasākumu (laboratoriski-diagnostiskie izmeklējumi, vakcinācijas u.c.) izpildi, kas, nodrošina dzīvnieku infekcijas slimību situācijas kontroli valstī. Tas rada priekšnosacījumus dzīvnieku infekcijas slimību riska prognozei un savlaicīgai slimību diagnosticēšanai.

**RĪCĪBAS PLĀNS** ir dokuments, kas nosaka dzīvnieku infekcijas slimību valsts uzraudzības augstākminēto pasākumu reglamentu, biežumu un metodes, kā priekšnoteikumu valsts pasargāšanai no dzīvnieku infekcijas slimību izplatības, kā arī kvalitatīvas un nekaitīgas pārtikas iegūšanai.

**RĪCĪBAS PLĀNS** ir izstrādāts un tiek realizēts saskaņā ar Veterinārmedicīnas likumu un attiecīgiem Ministru kabineta noteikumiem, tā izpildi kontrolē Pārtikas un veterinārais dienests.

**RĪCĪBAS PLĀNA PIELIKUMĀ Nr. 3** ir dots ieteicamo laboratoriski-diagnostisko izmeklējumu un profilaktisko vakcināciju saraksts, kurā minētie pasākumi ir būtiski atsevišķu dzīvnieku sugu veselības nodrošināšanai un kontrolei. Šo pasākumu izvēli un veikšanu nosaka dzīvnieka īpašnieks, konsultējoties ar praktizējošo veterinārārstu.

**SASTĀDĪTĀJI**

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Veterinārās uzraudzības departamenta direktors  
 Dzīvnieku infekcijas slimību uzraudzības daļas vadītāja  
 PVD ģenerāldirektora padomnieks  
 Veterinārās uzraudzības departamenta Risku vadības daļas vadītājs  
 Epizootiju uzraudzības sektora vadītājs  
 Epizootiju uzraudzības sektora vecākais eksperts  
 Epizootiju uzraudzības sektora vadītāja vietniece  
 Valsts Veterinārmedicīnas diagnostikas centra direktora vietniece  
 Valsts Veterinārmedicīnas diagnostikas centra Seroloģijas daļas vadītāja  
 Vecākā referente (datorsalikums)

**SATURS:**

A	Dzīvnieku infekcijas slimību valsts uzraudzības obligātie diagnostiskie izmeklējumi . . . . .	5
B	Dzīvnieku A un B grupas infekcijas slimību monitorings . . . . .	11
C	Dzīvnieku infekcijas slimību profilaktiskās vakcinācijas . . . . .	12

**Pielikumi:**

Nr.1	"Paraugi putnu salmonelozes (S.typhimurium un S.enteritidis) laboratoriskai diagnostikai" . . . . .	13
Nr.2	"Piena paraugu ņemšana dzīvnieku infekcijas slimību seroloģiskai diagnostikai" . . . . .	14
Nr.3	Rekomendācijas dzīvnieku īpašniekiem . . . . .	15

## I VISPĀRĪGIE JAUTĀJUMI UN SKAIDROJUMI

1. Par rīcības plāna A sadaļas izpildi ir atbildīgi dzīvnieku turētāji, kontroli nodrošina Pārtikas un veterinārais dienests.
2. Rīcības plāna B sadaļā paredzētos pienākumus veic Pārtikas un veterinārais dienests. Dzīvnieku infekcijas slimību (A un B grupu) epizootiskā fona noteikšanai paraugus ņem Pārtikas un veterinārā dienesta teritoriālās struktūrvienības valsts veterinārais inspektors; diagnostikumam un laboratorisko izmeklējumu izmaksas sedz no valsts budžeta līdzekļiem.
3. Rīcības plāna C sadaļā paredzētās vakcinācijas nodrošina (atbilstoši rīcības plāna pozīcijām) dzīvnieku turētāji un valsts.
4. Epidemioloģiskās izmeklēšanas procesā PVD inspektori ņemtos paraugus apmaksā no valsts budžeta līdzekļiem (epizootiju jeb A grupas slimību un zoonožu diagnostikai).
5. Ja plānā minēto izmeklējumu nav iespējams veikt dzīvnieka izcelsmes saimniecībā, tas jāveic saimniecībā, uz kuru dzīvnieks ir pārvietots, bet līdz izmeklējumu rezultātu iegūšanai dzīvniekam jāatrodas nošķirtam no pārējiem ganāmpulka dzīvniekiem, kas izslēgtu inficēšanas iespēju.
6. Ja saimniecībā konstatēti jebkādi pārkāpumi, kuru dēļ nevar izslēgt ganāmpulka inficēšanās iespējamību ar plānā minētajām slimībām, jāveic visu ganāmpulka dzīvnieku izmeklēšana.
7. Ja saimniecībā konstatē kādu no dzīvnieku infekcijas slimībām, tai tiek noteikti dzīvnieku pārvietošanas ierobežojumi un veikti pasākumi saskaņā ar normatīvo aktu prasībām.
8. Dzīvniekiem, kuri nav sasnieguši vecumu, kāds noteikts dzīvnieka izmeklēšanai uz konkrēto slimību, piemēro mātes izmeklējumu rezultātus.
9. Dzīvnieku aborta gadījumā piilvarotam veterinārārstam jāizvērtē iespējamie aborta cēloņi un, ja nevar izslēgt dzīvnieku saslimšanu ar kādu no zoonozēm, jānodrošina paraugus nosūtīšana laboratoriskai diagnostikai.
10. Rīcības plānā minēto diagnostisko izmeklējumu metodes un to saīsinājumi:
  - KSR – komplementa saistīšanas reakcija
  - IDR – imūndifūzijas reakcija
  - IFA – imūnfermatīvā analīze (ELISA)
  - IFA Bio-Rad Platella – ātrais tests GSE un Skrēpi laboratoriskai diagnostikai
  - Prionics WB – ātrais tests GSE un Skrēpi laboratoriskai diagnostikai
  - IFA DAKO tests – imūnfermatīvā analīze pasterellu toksīnu noteikšanai
  - HAAR – hemaglutinācijas aiztures reakcija
  - AR – aglutinācijas reakcija
  - VNR – vīrusa neitralizācijas reakcija (tests)
  - FAVN – fluorescējošo antivielu vīrusa neitralizācija
  - MAR – mikroaglutinācijas reakcija

**A DZĪVNIEKU INFEKCIJAS SLIMĪBU VALSTS  
 UZRAUDZĪBAS OBLIGĀTIE DIAGNOSTISKIE IZMEKLĒJUMI**

Nr. p. k.	Dzīvnieku suga, vecuma grupa, kategorija	Slimība, kods	Diagnostiskie izmeklējumi			Specifiskā profilakse	Piezīmes
			Biezums, apjoms	Metode	Apmaksa		
1	2	3	4	5	6	7	8
1.	<b>GOVIS</b>						
1.1.	<b>SERTIFICĒTI VAISLAS BUĻĻI</b>						
		1. Tuberkuloze /Tuberculosis/ B105	1 x gadā, 100% dzīvnieku	Alerģiska - tuberkulīna tests (zīdītāju tuberkulīns)	Tuberkulīns - no valsts budžeta, darba izmaksas sedz dzīvnieka īpašnieks	Vakcinācija aizliegta	
		2. Bruceloze /Brucellosis/ B103	1 x gadā, 100% dzīvnieku	Seroloģiska - rozbengāla reakcija (AR, KSR, IFA)	Visas izmaksas sedz dzīvnieka īpašnieks		
		3. Enzootiskā leikozā /Leucosis enzootica boum/ B108	1 x gadā, 100% dzīvnieku	Seroloģiska - IFA			
		4. Kampilobakterioze /Campylobacteriosis/ B104	1 x gadā, 100% dzīvnieku (spermas savākšanas periodā un/vai kuri ir kontakta ar buļļiem, no kuriem savāc spermu. Buļļi, pēc atpūtas perioda, kas garāks par 6 mēnešiem jāpārbauda ne vairāk kā 30 dienu laikā pirms plānotās spermas savākšanas uzsākšanas)	Mikrobioloģiska - prepūcija gļotās vai sekretos, spermā		Specifiskas profilakses nav	
		5. Trihomonozē /Trichomonosis/ B112	1 x gadā, 100% dzīvnieku (spermas iegūšanas periodā un/vai kuri ir kontakta ar buļļiem, no kuriem savāc spermu. Buļļi, pēc atpūtas perioda, kas garāks par 6 mēnešiem jāpārbauda 30 dienu laikā pirms plānotās spermas savākšanas uzsākšanas)	Mikrobioloģiska - prepūcija gļotās vai sekretos, spermā		Specifiskas profilakses nav	
		6. Govju virusālā diareja /Bovine virus diarrhoea/ C652	1 x gadā, 100% dzīvnieku	Seroloģiska - IFA, nosakot antivielas. Ja buļļis ir seropozitīvs - jāpārbauda uz vīrusa klātbūtni (vai jāiznīcina) katrs iegūtais ejakulāts kopš pēdēja negatīvā seroloģiskā testa.		Maksimālās apsekošanas staciju un punktu dzīvniekiem profilaktiska vakcinācija aizliegta	

**PĀRTIKAS UN VETERINĀRAIS DIENESTS**

**DZĪVNIĒKU INFEKCIJAS SLIMĪBU VALSTS UZRAUDZĪBAS 2004. GADA RĪCĪBAS PLĀNS**

1	2	3	4	5	6	7	8
		7. Infekciozais rinotraheīts-pustulozais vulvovaginīts (IRT-PVV) /Rhinotracheitis infectiosa boum-vulvovaginitis pustulosa/ B110	1 x gadā, 100% dzīvnieku	Seroloģiska - IFA, nosakot antivielas			
		8. Hlamīdioze /Chlamydiosis/	1 x gadā, 100% dzīvnieku	Seroloģiska – KSR vai IFA	Visas izmaksas sedz dzīvnieka īpašnieks	Maksimālās apsekošanas staciju un punktu dzīvniekiem profilaktiska vakcinācija aizliegta	
		9. Paratuberkuloze /Paratuberculosis/ B059	1 x gadā, 100% dzīvnieku	Seroloģiska - IFA		Specifiskās profilakses nav	
						Vakcinācija aizliegta	
<b>1.2.</b>	<b>GOVJU SUGAS DZĪVNIĒKI</b>						
	Govju sugas dzīvnieki no 8 nedēļu vecuma	1. Tuberkuloze /Tuberculosis/ B105	1 x gadā, 100% dzīvnieku, izņemot saimniecības, kurās govīs netiek turētas kopā ar citu sugu dzīvniekiem - izmeklējumus veic katru otro gadu	Alerģiska - tuberkulīna tests (zīdītāju tuberkulīns)	Tuberkulīns – no valsts budžeta, darba izmaksas sedz dzīvnieka īpašnieks		
	Govju sugas dzīvnieki no 12 mēnešu vecuma, izņemot nobarojamās vīrišu kārtas dzīvniekus	2. Bruceloze /Brucellosis/ B103	katru otro gadu 100% dzīvnieku	Seroloģiska - rozbengāla reakcija (AR, KSR, IFA) vai piena paraugā ar IFA		Vakcinācija aizliegta	
	Govju sugas dzīvnieki no 24 mēnešu vecuma	3. Enzootiskā leiکوze /Leucosis enzootica boum/ B108	a) katru otro gadu 100% dzīvnieku (Aizkraukles, Jelkabpils, Jelgavas, Liepājas, Ogres, Saldus, Talsu, Tukuma, Valkas, Valmieras, Ventspils rajonos) b) 1 x gadā, 100% dzīvnieku (Alūksnes, Balvu, Bauskas, Cēsu, Daugavpils, Dobeles, Gulbenes, Krāslavas, Kuldīgas, Limbažu, Ludzas, Madonas, Preiļu, Rēzeknes, Rīgas)	Seroloģiska - IFA vai piena paraugā ar IFA	Visas izmaksas sedz dzīvnieka īpašnieks		Ja piena paraugi ir pozitīvi, jāveic individuālā seroloģiska dzīvnieku izmeklēšana

- Pārvietojot govju sugas dzīvniekus uz citu ganāmpulku, 30 dienu laikā pirms pārvietošanas tie jāizmeklē uz tuberkulozi, brucelozi un leiکوzi, ja 2004. gadā ganāmpulka dzīvnieki nav izmeklēti uz minētajām slimībām. Pārvietošanas gadījumā jāizmeklē uz leiکوzi no 12 mēnešu vecuma.
- Nobarojamiem dzīvniekiem piemēro mātes izmeklējumu rezultātus.

PĀRTIKAS UN VETERINĀRAIS DIENESTS

DZĪVIEKŪ INFEKCIJAS SLIMĪBU VALSTS UZRAUDZĪBAS 2004. GADA RĪCĪBAS PLĀNS

1	2	3	4	5	6	7	8	
2.	<b>CŪKAS</b>							
2.1.	<b>VAISLAS KUIĻI, KURUS IZMANTO MĀKSLĪGĀS APSĒKĻOŠANAS STACIJĀS UN PUNKTOS</b>							
		1. Klasiskais cūku mēris /Pestis suum/ A130	1 x gadā, 100% dzīvnieku	Seroloģiska – IFA nosakot antivielas	Laboratoriskās diagnostikas izmaksas sedz no valsts budžeta			
		2. Bruceloze /Brucellosis/ (ferosinātais B.suis) B253	1 x gadā, 100% dzīvnieku	Seroloģiska - rozbengala reakcija (KSR)	Visas izmaksas sedz dzīvnieka īpašnieks			
		3. Aujeski slimība /Morbus Aujesky/ B052	1 x gadā, 100% dzīvnieku	Seroloģiska - IFA				
		4. Tuberkuloze /Tuberculosis/	1 x gadā, 100% dzīvnieku	Alerģiska - tuberkulīna tests (ar zīdītāju un putnu tuberkulīnu)	Tuberkulīns – no valsts budžeta, darba izmaksas sedz dzīvnieka īpašnieks	Vakcinācija aizliegta		
		5. Cūku infekciozais atroflakais rinīts /Rhinitis infectiosa atrophica suum/ B251	1 x gadā, 100% dzīvnieku	Mikrobioloģiska, izdalot pastereļas un seroloģiska - IFA DAKO tests pastereļļu toksīnu noteikšanai				
		6. Cūku reprodūktīvais respiratorais sindroms B257	1 x gadā, 100% dzīvnieku	Seroloģiska - IFA				
		7. Cūku parvovīrusālā infekcija	1 x gadā, 100% dzīvnieku	Seroloģiska - HAAR vai IFA	Visas izmaksas sedz dzīvnieka īpašnieks	Vakcinācija ieteicama	Vakcinātos kuliņus neizmeklē	
		8. Leptospiroze / Leptospirosis / B056	1 x gadā, 100% dzīvnieku	Seroloģiska - MAR		Vakcinācija ar attiecīgā serotipa vakcīnu atļauta		
2.2.	<b>SIVĒNMĀTES, REMONTCŪKAS, VAISLAS KUIĻI, KURUS IZMANTO LECINĀŠANAI (APSĒKĻOŠANAI) SAVĀ GANĀMPULKĀ</b>							
	Sivēnmātes, vaislas kuliņi un remontcūkas	1.Tuberkuloze /Tuberculosis/	1 x gadā: 10% dzīvnieku ganāmpulkā (bet ne mazāk kā 10); ja ganāmpulkā 1-10 dzīvnieki - izmeklē visus	Alerģiska - tuberkulīna tests (ar zīdītāju un putnu tuberkulīnu)	Tuberkulīns – no valsts budžeta, darba izmaksas sedz dzīvnieka īpašnieks	Vakcinācija aizliegta		
	Sivēnmātes, vaislas kuliņi un remontcūkas	2.Bruceloze /Brucellosis/ (ferosinātais - B.suis) B253	1 x gadā: 10% dzīvnieku ganāmpulkā (bet ne mazāk kā 10); ja ganāmpulkā 1-10 dzīvnieki - izmeklē visus	Seroloģiska - rozbengala reakcija				
	Sivēnmātes un vaislas kuliņi Remontcūkas	3.Aujeski slimība /Morbus Aujesky/ B052	1 x gadā: 10% dzīvnieku ganāmpulkā (bet ne mazāk kā 10), ja ganāmpulkā 1-10 dzīvnieki - izmeklē visus Pirms lecīnāšanas vai sēkļošanas, vai pirms ievietošanas ganāmpulkā - 100% dzīvnieku, ja ganāmpulks pakļauts slimības apkārošanas programmai	Seroloģiska – IFA	Visas izmaksas sedz dzīvnieka īpašnieks	Vakcinācija ar iezīmētu vakcīnu		

**PĀRTIKAS UN VETERINĀRAIS DIENESTS**  
**DZĪVNIĒKU INFEKCIJAS SLIMĪBU VALSTS UZRAUDZĪBAS 2004. GADA RĪCĪBAS PLĀNS**

1	2	3	4	5	6	7	8
<b>3. ZIRGI</b>							
<b>3.1. SERTIFICĒTI VAISLAS ĒRZELI</b>							
	1. Zirgu jaunie ienāši /Malleus/ B209	1 x gadā, 100% dzīvnieku	Seroloģiska - KSR	Visas izmaksas sedz dzīvnieka īpašnieks			
	2. Infekciāzā anēmija /Anaemia infectiosa equorum/ B205	1 x gadā, 100% dzīvnieku	Seroloģiska - IDR				
	3. Infekciāzais arterīts /Arteritis viralis equorum/ B211	1 x gadā, 100% dzīvnieku, uzsākot lecināšanas (apsēklošanas) sezonu	Seroloģiska - VNR	Laboratoriskos izmeklējumus apmaksā no valsts budžeta līdzekļiem			
<b>3.2. ZIRGI</b>							
	1. Infekciāzā anēmija /Anaemia infectiosa equorum/ B205	1 x gadā, 100% dzīvnieku	Seroloģiska - IDR	Visas izmaksas sedz dzīvnieka īpašnieks			
	2. Zirgu jaunie ienāši /Malleus/ B209	1 x gadā, 100% dzīvnieku	Seroloģiska - KSR				
<b>4. AITAS</b>							
<b>4.1. SERTIFICĒTI VAISLAS TEĶI</b>							
	1. Teķu infekciāzais epididimīts /Epididymitis infectiosa arietum/ (ierosinātājs - B.ovis) B151	1 x gadā, 100% dzīvnieku	Seroloģiska - KSR	Visas izmaksas sedz dzīvnieka īpašnieks	Vakcinācija aizliegta		
	2. Listerioze /Listeriosis/ C611	1 x gadā, 100% dzīvnieku	Seroloģiska - KSR		Vakcinācija atļauta	Vakcinētos dzīvniekus neizmeklē	
	3. Maedi - Visna slimība B161	1 x gadā, 100% dzīvnieku	Seroloģiska - IFA		Specifiskās profilakses nav		
<b>4.2. AITAS UN VAISLAS TEĶI, KURUS IZMANTO LECINĀŠANAI (APSĒKLOŠANAI) SAVĀ GANĀMPULKĀ</b>							
No 6 mēnešu vecuma	1. Teķu infekciāzais epididimīts /Epididymitis infectiosa arietum/ (ierosinātājs B.ovis) B151	1x gadā, 100% vaislas teķu	Seroloģiska - KSR	Visas izmaksas sedz dzīvnieka īpašnieks	Vakcinācija aizliegta		
	2. Listerioze /Listeriosis/ C611	1x gadā 10% dzīvnieku ganāmpulkā (bet ne mazāk kā 10); ja ganāmpulkā 1-10 dzīvnieku, izmeklē visus	Seroloģiska - KSR		Vakcinācija atļauta	Vakcinētos dzīvniekus neizmeklē	

PĀRTIKAS UN VETERINĀRAIS DIENESTS

DZĪVNIĒKU INFEKCIJAS SLIMĪBU VALSTS UZRAUDZĪBAS 2004. GADA RĪCĪBAS PLĀNS

1	2	3	4	5	6	7	8
5.	<b>KAZAS</b>						
5.1.	<b>SERTIFICĒTI VAISLAS ĀŽI</b>						
		1. Kazu artrīts-encefalīts /Caprine arthritis encephalitis/ B153	1 x gadā, 100% dzīvnieku	Seroloģiska - IFA	Visas izmaksas sedz dzīvnieka īpašnieks	Specifiskās profilakses nav	
6.	<b>PUTNI</b>						
	Vaislas putnu ganāmpulkos un ganāmpulkos, no kuriem iegūst pārtikas olas	1. Salmoneloze /Salmonellosis/ 1.1. S.pulorum un S.gallinarum vistām, tītariem, pīļiem, pērļu vistipām, paipalām, lrbēm, fazāniem 1.2. S.arizonae tītariem	Izmeklē katrā dešanas cikla laikā, ne mazāk kā 10% no vaislas putnu ganāmpulka, pārtikas olu ražošanas saimniecībās: ja 1-20 putni - 100%; 20 līdz 100 - ne mazāk par 20 putniem; 100 līdz 1000 - 10%, bet ne mazāk par 20 putniem; ja vairāk par 1000 - 1%, bet ne mazāk kā 100 putnus. Inkubatorā profilaktisko kontroli veic pirms katras olu partijas ievietošanas, bet cāļu audzēšanas telpās - pirms cāļu ievietošanas.	Seroloģiska un /vai mikrobioloģiska Paraugi: asinis, braketie cāļi, dūnas vai putekļi no inkubatora, uztriepe no inkubatora sienām, pakāisi vai ūdens paraugi no dzirdnēm	Diagnostikums (pulorozes antigēns) - no valsts budžeta, darba, I.sk. laboratorijas izmaksas sedz dzīvnieka īpašnieks		
	Vaislas putnu ganāmpulkos, ganāmpulkos, no kuriem iegūst pārtikas olas un broileru ganāmpulkos	1.3. S.typhimurium, 1.4. S.enteritidis	skat. Pielikumu Nr.1		Paškontroles paraugu izmeklēšanu apmaksā īpašnieks, valsts veterinārā inspektoraņemto oficiālo paraugu izmeklēšanu apmaksā no valsts budžeta līdzekļiem	Specifiskās profilakses nav	
	Vaislas vistu un tītaru ganāmpulkos	2. Mikoplazmoze /Mycoplasmosis/ B311 2.1. M.gallisepticum	Paraugus ņem audzēšanas un dešanas laikā, tajā skaitā tieši pirms dešanas uzsākšanas un turpmāk ik pēc 3 mēnešiem. Izmeklē 5% no ganāmpulka, bet ne vairāk kā 100 putnus.	Seroloģiska - AR, (IFA, HAAR) un/vai mikrobioloģiska. Paraugi: asinis, dienu veci cāļi un tītareni, sperma, uztriepes no trahejas, kloakas vai gaisa maisiem	Visas izmaksas sedz dzīvnieka īpašnieks		
	Vaislas tītaru ganāmpulkos	2.2. M.meleagridis					

- Saimniecībām, kuras nodarbojas ar putnu audzēšanu, jāizstrādā putnu salmonelozes un mikoplazmozes paškontroles programmas.

- Ik pēc 8 nedēļām PVD veterinārais inspektors vaislas putnu novietnē ņem oficiālu paraugu. Šajos gadījumos paškontroles paraugi nav jāņem.

**PARTIKAS UN VETERINĀRAIS DIENESTS**  
**DZĪVNIKU INFEKCIJAS SLIMĪBU VALSTS UZRAUDZĪBAS 2004. GADA RĪCĪBAS PLĀNS**

1	2	3	4	5	6	7	8
7.	<b>ZIVIS ZIVJAUDZĒTAVĀS</b>						
	Zivis (lašveidīgās, līdakas, ālatas)	1. Virālā hemorāģiska septicēmija /Viral haemorrhagic septicaemia/ B401	1 x gadā klīniski un laboratoriski	Klīniska, patoloģiskā un virusoloģiska - virusa izolešana un identificēšana audu kultūrās ar VNR	Visas izmaksas sedz īpašnieks		Laboratoriskie izmeklējumi jāuzsāk saskaņā ar atsevišķu PVD rīkojumu
	Zivis (lašveidīgās)	2. Infekciozā hematopoētiskā nekroze /infectious haemopoietic necrosis/ B405					
	Zivis (lašveidīgās)	3. Lašveidīgo infekciozā anēmija /Anaemia infectious salmonidae/					
Zivis (karpas)	4. Karpu sarkanā sērga						
8.	<b>VĒŽI VĒŽU AUDZĒTAVĀS</b>						
		1. Vēžu baltplankumu slimība	1 x gadā	Mikoloģiska	Visas izmaksas sedz īpašnieks	Specifiskās profilakses nav	
		2. Vēžu mēris /Pestis astaci/	1 x gadā				
9.	<b>BĪŠU</b>						
	Bišu saimes	1. Eiropas peru puve / European foulbrood / B453	2 x gadā (pavasari un rudenī)	Klīniska	Visas izmaksas sedz bišu dravas īpašnieks	Specifiskās profilakses nav	
	Bišu saimes, kurās nobeigušies perē		Saskaņā ar praktizējošā veterinārārsta norādījumiem diagnozes noskaidrošanai	Mikrobioloģiska			
	Bišu saimes	2. Amerikas peru puve / American foulbrood / B452	2 x gadā (pavasari un rudenī)	Klīniska			
Bišu saimes, kurās nobeigušies perē	Saskaņā ar praktizējošā veterinārārsta norādījumiem diagnozes noskaidrošanai		Mikrobioloģiska				

## B DZĪVNIĒKU A UN B GRUPAS INFEKCIJAS SLIMĪBU EPIZOOTISKĀ FONĀ NOTEIKŠANA

Nr. p.k.	Slimība, kods	Dzīvnieki	Izmeklējamā substance	Izmeklējamo dzīvnieku kopskaits*	Izmeklējuma metode
1	2	3	4	5	6
1.	Klasiskais cūku mēris /Pestis suum/ A130	Cūkas	Asins seruma paraugi	1000	IFA, pozitīviem -VNR
		Nomedītās savvaļas cūkas	Parenhimatozo orgānu paraugi un asins seruma paraugi	520	IFA uz antigēnu un antivielām
2.	Putnu gripa /Grippus avium/ A150	Putni	Asins seruma paraugi	800	HAAR vai IFA
3.	Nokāslas slimība /Morbus newcastle/ A160	Putni	Putnu smadzeņu, trahejas un plaušu, aknu paraugi (ņemti izlases veidā putnu kaušanas procesā)	300	Vīrusa izolēšana un tā intracerebrālās patogenitātes indeksa noteikšana
4.	Govju sūkņveida encefalopātija (GSE) / Encephalopathiae spongiformium boum/ B115/	Govis	Galvas smadzeņu audu paraugi no: a)visām kritušām govīm, vecākām par 24 mēnešiem; b)visām piespiedu kārtā nokautajām govīm, vecākām par 24 mēnešiem; c)visām govīm, vecākām par 24 mēnešiem, kurām pirms kaušanas apskates laikā konstatētas klīniskās pazīmes; d)veselām, cilvēku patēriņam kautām govīm, vecākām par 30 mēnešiem (saskaņā ar atsevišķu PVD rīkojumu), e)visu vecumu klīniski uz GSE aizdomīgiem govju sugas dzīvniekiem; f)visiem dzīvniekiem, kuri nogalināti GSE apkaršanas laikā	-	IFA Bio-Rad Platēla, Prionics WB, histoloģiska, imūnhistokīmiska
5.	Skrepi /Chesmus ovium/ B160	Aitas, kazas	Galvas smadzeņu audu paraugi no: a)visām aitām/kazām, kritušām vai nogalinātām, vecākām par 18 mēnešiem vai kurām nomainījušies priekšzobi; b)visu vecumu uz skrepi klīniski aizdomīgām aitām un kazām, c)visiem dzīvniekiem, kuri nogalināti skrepi apkaršanas laikā	-	IFA Bio-Rad Platēla, Prionics WB, histoloģiska, imūnhistokīmiska
6.	Trakumsērga /Rabies/ B058	Nomedītās lapsas	Apakšzokļa kauls	600	Tetraciklīna noteikšana ar luminescences mikroskopu kaulaudos
			Asins seruma paraugi	150	Seroloģiski - FAVN
7.	Bruceloze /Brucellosis/ (ierosinātais - B.melitensis) B152	Aitas, kazas	Asins seruma paraugi	1800 aitas, 700 kazas	Seroloģiska - rozbengāla reakcija (AR, KSR, IFA)

\* Izmeklējamo dzīvnieku skaits atsevišķos rajonos tiek noteikts ar atsevišķu PVD rīkojumu.



1. pielikums

## PARAUGI PUTNU SALMONELOZES (S.TYPHIMURIUM, S. ENTERITIDIS) LABORATORISKAJAI DIAGNOSTIKAI

### I Vaislai audzējamo putnu ganāmpulkos paraugus ņem:

- 1 dienu veciem cāļiem\*:
  - noskalojumus no kontaineru iekšējām virsmām, kuros cāļi piegādāti uz novietni;
  - izmeklējamu materiālu no cāļiem, kas nobeigušies pārvešanas laikā;
2. 4 nedēļas veciem jaunputniem:
  - fekāliju kopparaugus;
3. jaunputniem 2 nedēļas pirms dēšanas cikla uzsākšanas:
  - fekāliju kopparaugus.

### Fekāliju kopparaugi\*\* atbilstoši putnu skaitam novietnē

Putnu skaits novietnē	Vietu skaits putnu novietnē, no kurām tiek ņemti fekāliju paraugi tikpat, cik putni, bet ne vairāk kā 20
1-24	
25-29	20
30-39	25
40-49	30
50-59	35
60-89	40
90-199	50
200-499	55
500 un >	60

\* cāļi, kas jaunāki par 72 stundām un nav saņēmuši barību

\*\* fekāliju kopparaugu veido no dažādās novietnes vietās ņemtiem atsevišķiem svaigu fekāliju paraugiem, kas katrs sver ne mazāk kā 1 gramu.

### II Pieaugušu vaislas putnu ganāmpulkos paraugus ņem:

1. Vismaz vienu reizi divās nedēļās:
  - a) ganāmpulkos, no kuriem putnu olas tiek inkubētas inkubatorā, kura kapacitāte ir līdz 1000 olām:
    - fekāliju kopparaugus;
  - b) ganāmpulkos, no kuriem putnu olas tiek inkubētas inkubatorā, kura kapacitāte ir 1000 un vairāk olas, jāzmeklē inkubatorā ņemot šādu paraugu:
    - 250 izšķīlušos cāļiņu (olas inkubatorā iesūtītas no viena ganāmpulka) mekonija kopparaugu vai,
    - izmeklējamais materiāls no 50 cāļiņiem, kuri nobeigušies olas (olas inkubatorā iesūtītas no viena ganāmpulka) inkubācijas laikā.
2. Šādi paraugi jāņem arī vaislas putnu ganāmpulkos, kuros ir mazāk par 250 putniem, kuru olas tiek nosūtītas uz inkubatoru, kura kapacitāte ir 1000 un vairāk olas.
3. Katru astoto nedēļu II punktā minēto oficiālo paraugu ņemšanu veic PVD inspektors.

### III Putnu ganāmpulkos, no kuriem iegūst pārtikas olas, paraugus ņem:

1. 1 dienu veciem cāļiem (kā I.1. punktā);
2. jaunputniem 2 nedēļas pirms dēšanas cikla uzsākšanas – fekāliju kopparaugus;
3. dēšanas sezonas laikā ņem fekāliju kopparaugus trīs reizes – 30 nedēļu vecumā, 50 nedēļu vecumā un 4 nedēļas pirms nosūtīšanas uz kautuvi.

### IV Broilieru audzētāju saimniecībās paraugus ņem:

1. 1 dienu veciem cāļiem (kā punktā I.1.);
2. broilieriem nedēļu pirms nosūtīšanas uz kautuvi – fekāliju kopparaugu vai kopējo paraugu, izgriežot kļeakas 10 putnu līķiem

## 2. pielikums **GOVJU PIENA PARAUGU NOŅEMŠANA DZĪVNIKU INFEKCIJAS SLIMĪBU SEROLOĢISKAI DIAGNOSTIKAI**

**Piena paraugu iegūšanā, komplektēšanā, iepakojšanā un nostādīšanā uz veterinārajām laboratorijām jāievēro vairāki nosacījumi!**

1. piena paraugi veterinārajās laboratorijās jāiesūta tīros, ķīmiski neitrālos, vienreizējas lietošanas, apmēram 30 ml tilpuma plastmasas trauciņos vai stobriņos, kas sertificēti bioloģisku produktu iepakojšanai, ar skrūvējamu vai blīvi aizspiežamu vāciņu (piemēram, piena pārraudzības izmekļojumiem izmantojamie trauciņi);
2. piena paraugus izmeklēšanai uz govju enzootisko leikozi iesūta tikai no leikozes brīviem ganāmpulkiem;
3. veterinārajās laboratorijās var iesūtīt 3 veida piena paraugus (paraugu veidu pareizi nosaucot pavaddokumentāl):
  - > **individuālu piena paraugu** - piena paraugu no vienas laktējošas govys,
  - > **fermā apvienotu piena paraugu** - paraugu, kas iegūts fermā apvienojot līdz 50 (!) laktējošu govju vienāda apjoma (!) piena paraugus, lai nepārsniegtu diagnostikai izmantojamo reagentu jutības indeksu (t.i., 1:50). Paraugs jāgatavo ļoti precīzi - no katra dzīvnieka jāņem vienāds piena parauga daudzums, iegūtais apvienotais piena paraugs rūpīgi jāsamaisa un uz laboratoriju jānosūta ne vairāk kā 30 ml apvienotā piena,
  - > **kopplena paraugu no piena uzglabāšanas sistēmas** - saimniecībās, kurās ir līdz 10 laktējošām govīm;

4. paraugi laboratorijās jāiesūta ne vēlāk kā 48 stundu laikā, uzglabājot no +2 līdz +8°C temperatūrā. Ja šādu uzglabāšanas režīmu nav iespējams nodrošināt, tad jālieto konservants (konservanta aktīvā viela - 2-brom-2-nitropropān-1,3-diols) saskaņā ar konservanta lietošanas instrukciju (ja paraugi konservēti, tas jānorāda pavadrakstā!);
5. slaukšanas reizei (rīta, pusdienas vai vakara) nav ietekme uz antivielu līmeni pienā;
6. pirms parauga noņemšanas noslauc pirmās piena strūkļas;
7. piena paraugi izmeklēšanai uz leikozi u.c. infekcijas slimībām nav derīgi no govīm 2 nedēļas pirms ciellaišanas un 2 nedēļas pēc atnešanās, kā arī no govīm ar piena dziedzeru iekaisumiem.

Ja piena paraugam iegūts pozitīvs vai aizdomīgs rezultāts, atkārtotai izmeklēšanai jāiesūta asins paraugs (ja pozitīvs vai aizdomīgs rezultāts iegūts fermā apvienotajam piena paraugam, tad jāiesūta asins paraugi no katra šādas grupas dzīvnieka).

3. pielikums

## REKOMENDĀCIJAS DZĪVNIEKU ĪPAŠNIEKIEM / ieteicamie diagnostiskie izmeklējumi atsevišķu sugu dzīvniekiem /

N. p.k.	Dzīvnieku suga	Diagnostiskie izmeklējumi uz:
1.	Govis	- Paratuberkulozi - Govju virusālo diareju - Infekciozo rinotraheītu – pustulozo vulvovaginītu - Parazītu klātbūtni
2.	Cūkas	- Cūku reproductīvo respiratoro sindromu - Cūku infekciozo atrofisko rinītu - Tuberkulozi - Mikoplazmozi - Parazītu klātbūtni
3.	Zirgi	- Rīnopneimoniju - Zirgu virālo arterītu - Parazītu klātbūtni
4.	Aitas	- Maedi-Visna slimību - Parazītu klātbūtni
5.	Kazas	- Kazu artrītu-encefalītu - Parazītu klātbūtni
6.	Putni	- Tuberkulozi - Parazītu klātbūtni
7.	Zivis	- Pavasara karpu virēmiju - Parazītu klātbūtni
8.	Bites	- Akarapidozi - Nozematozi - Varrozi
9.	Kazokzvēri	- Trihinelozi (seroloģiski) - Ūdeņu virusālo enterītu - Parazītu klātbūtni
10.	Kaķi	- Toksoplazmoze
11.	Suņi	- Toksoplazmoze

## REKOMENDĀCIJAS DZĪVNIKU ĪPAŠNIEKIEM

/ ieteicamās profilaktiskās vakcinācijas atsevišķu sugu dzīvniekiem /

N. p.k.	Dzīvnieku suga	Profilaktiskās vakcinācijas pret
1.	Govis	- Infekciozo rinotraheītu – pustulozo vulvovaginītu ganāmpulkos, kuros izdala seroloģiski pozitīvus dzīvniekus - Kailo ēdi
2.	Cūkas	- Aujeski slimību ganāmpulkos, kuros izdala seroloģiski pozitīvus dzīvniekus - Cūku reproductīvo respiratoro sindromu ganāmpulkos, kuros izdala seroloģiski pozitīvus dzīvniekus - Cūku parvovirusālo infekciju - Cūku infekciozo atrofisko rinītu ganāmpulkos, kuros izdala seroloģiski pozitīvus dzīvniekus - Cūku sarkangūļu
3.	Zirgi	- Rinopneimoniju - Tetanus - Zirgu gripu - Kailo ēdi
4.	Putni	- Nākšas slimību - Gamboro slimību - Mareka slimību - Infekciozo bronhītu - Reovīrusa infekciju
5.	Truši	- Miksomatozi - Trušu hemorāģisko slimību
6.	Kažokzvēri	- Gaļedāju mēri - Ūdeņu virusālo enterītu - Botulismu - Salmonelozi
7.	Kaķi	- Kalicivīrusālo infekciju - Leptospirozi - Rinotraheītu - Panleikopēniju
8.	Suņi	- Parvovirozi - Leptospirozi - Gaļedāju mēri - Infekciozo hepatītu

**Annual Report****FAO/OIE/WHO Questionnaire - 2003  
Latvia****Head of Veterinary Services**

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**VETERINARIANS AND TECHNICAL PERSONNEL**

ACTIVITY	NUMBER
VETERINARIANS Government officials (central, local)	353
In laboratories, universities, training institutions	100
Private practitioners	870
Other (please specify in note below)	0
<b>TOTAL =</b>	<b>1323</b>

TECHNICAL Animal health assistants (with formal training)	121
PERSONNEL Animal health auxiliaries	21
Involved in food hygiene, including meat inspectors	0
<b>TOTAL =</b>	<b>142</b>

**FARMED ANIMALS**

ANIMAL GROUP	LIVESTOCK POPULATION	NUMBER OF ESTABLISHMENTS
cattle	389639	80336
buffaloes	0	0
sheep	39200	3600

goats	15000	13900
equidae	15400	1500
camelidae	0	0
cervidae	0	0
swine	444400	21500
birds	4002600	77900
rabbits/hares	149184	12900

ANIMAL GROUP	PRODUCTION (kg)	NUMBER OF AQUACULTURE ESTABLISHMENTS
fish	580675	154
molluscs	0	0
crustaceans	2000	26

ANIMAL GROUP	NUMBER OF HIVES	NUMBER OF APIARIES
bees	54148	6995

NOTE

**ANIMAL HEALTH AND CONTROL MEASURES  
LIST A DISEASES****A010 Foot and mouth disease Occurrence : 11/1987**

Outbreaks	Cases	Deaths	Control measures	Destroyed	Slaughtered	Vaccinated
bov			* / Qf / M / Vp			
	Note : 297 blood samples were tested					
sui			* / Qf / M / Vp			
	Note : 250 blood samples were tested					
Incidence Total = 0						



Note : 40 blood samples were tested  
ovi \* / Qf / M  
Note : 68 blood samples were tested  
Incidence Total = 0

**A110 African horse sickness** Occurrence : 0000  
Outbreaks Cases Deaths Control Destroyed Slaughtered Vaccinated  
measures \* / Qf / M  
equ  
Note : 50 blood samples were tested  
Incidence Total = 0

**A120 African swine fever** Occurrence : 0000  
Outbreaks Cases Deaths Control Destroyed Slaughtered Vaccinated  
measures \* / Qf / M  
sui  
Note : 94 blood samples were tested  
Incidence Total = 0

**A130 Classical swine fever** Occurrence : 04 / 1996  
Outbreaks Cases Deaths Control Destroyed Slaughtered Vaccinated  
measures \* / Qf / M /  
fau Vp  
Note : 355 samples were tested  
sui \* / Qf / M /  
Vp  
Note : 1217 blood samples were tested  
Incidence Total = 0

**A150 Highly pathogenic avian influenza** Occurrence : 0000  
Outbreaks Cases Deaths Control Destroyed Slaughtered Vaccinated  
measures \* / Qf / M /  
avi Vp

Note : 811 blood samples were tested  
Incidence Total = 0

**A160 Newcastle disease** Occurrence : 0000  
Outbreaks Cases Deaths Control Destroyed Slaughtered Vaccinated  
measures \* / Qf / M /  
avi V  
Note : 145 samples were tested  
Incidence Total = 0

**ANIMAL HEALTH AND CONTROL MEASURES**  
**LIST B DISEASES**

**B051 Anthrax** Occurrence : 01 / 1989  
Outbreaks Cases Deaths Control Destroyed Slaughtered Vaccinated  
measures \* / Qf / V  
bov  
Note : Animals vaccination against Anthrax carried out in stationary  
affected points 157  
cap \* / Qf / V  
Note : Animals vaccination against Anthrax carried out in stationary  
affected points 22  
equ \* / Qf / V  
Note : Animals vaccination against Anthrax carried out in stationary  
affected points 52  
ovi \* / Qf / V  
Note : Animals vaccination against Anthrax carried out in stationary  
affected points 9  
sui \* / Qf / V  
Note : Animals vaccination against Anthrax carried out in stationary  
affected points 127

**B052 Aujeszky's disease** Occurrence : 03 / 2002  
Outbreaks Cases Deaths Control Destroyed Slaughtered Vaccinated  
measures

sui \*/Te/  
Qf/V  
80304

Note : 9143 pigs were subjected to serological tests and 435 were positive, but were no clinical signs of disease

**B053 Echinococcosis/hydatidosis Occurrence : +?**

Outbreaks Cases Deaths Control Destroyed Slaughtered Vaccinated  
measures  
bov \*/Su

Note : out of 109495 cattle slaughtered, didn't find any infected

Note : out of 2775 ovis slaughtered and didn't find any infected

Note : out of 428581 swine slaughtered 1627 found infected

**B055 Heartwater Occurrence : 0000**

Outbreaks Cases Deaths Control Destroyed Slaughtered Vaccinated  
measures  
\*\*\* \*

**B056 Leptospirosis Occurrence : +?**

Outbreaks Cases Deaths Control Destroyed Slaughtered Vaccinated  
measures  
bov \*/Te/V 15

Note : 3400 blood samples were tested and 33 were positive, but there were no clinical signs of disease

Note : 76 blood samples were tested and 1 were positive, but there were no clinical signs of disease

Note : 69 blood samples were tested and 1 were positive, but there were no clinical signs of disease

Note : 18 blood samples were tested and all samples were negative

Note : 1056 blood samples were tested and 39 were positive, but there were no clinical signs of disease

**B057 Q fever Occurrence : 0000**

Outbreaks Cases Deaths Control Destroyed Slaughtered Vaccinated  
measures  
\*\*\* \*

**B058 Rabies Occurrence : +**

Outbreaks Cases Deaths Control Destroyed Slaughtered Vaccinated  
measures  
bov 20 21 14 Sp/\*/  
Qf/Qi/V/Z 7 4229

can 59 62 32 Sp/\*/  
Qf/Qi/V/Z 30 110824

cap 0 0 0 \*/Qf/V 68

equ 0 0 0 \*/Qf/V 95

fau 805 829 139 Sp/Cr/  
\*/Qi/V/Z 690 300000

Note : oral vaccination of wild foxes was performed in all country with 300 000 doses vaccine

fel 50 52 19 Sp/\*/  
Qf/Qi/V/Z 33 30446

ovi 0 0 0 \*/Qf/V 161

sui 0 0 0 \*/Qf/V 242

**B059 Paratuberculosis Occurrence : 10/1999**

Outbreaks Cases Deaths Control Destroyed Slaughtered Vaccinated  
measures  
bov \*/Te/  
Su/Qf/Qi

Note : 7061 blood samples were tested and 31 were positive, but there were no clinical signs of disease

cap \*/Qf

**B060 New world screwworm (C. hominivorax) Occurrence : 0000**

**B061 Old world screwworm  
(C. bezziana)** Occurrence : 0000

**B062 Trichinellosis** Occurrence : 03/2001

Outbreaks Cases Deaths Control Destroyed Slaughtered Vaccinated  
faun measures  
\*/Su/M  
sui \*/Te/M

Note : All slaughtered pigs are tested before entering the food chain.  
564 hunted wild boars and 43 foxes were tested, 3.2% wild boars and  
48,8% foxes were positive

**B101 Bovine anaplasmosis** Occurrence : 0000

Outbreaks Cases Deaths Control Destroyed Slaughtered Vaccinated  
bov measures \*

**B102 Bovine babesiosis** Occurrence : 09/1988

Outbreaks Cases Deaths Control Destroyed Slaughtered Vaccinated  
bov measures \*

**B103 Bovine brucellosis** Occurrence : 1963

Outbreaks Cases Deaths Control Destroyed Slaughtered Vaccinated  
bov measures \*/Te/qf

Note : 208135 blood samples were tested with negative results

**B104 Bovine genital  
campylobacteriosis** Occurrence : 0000

Outbreaks Cases Deaths Control Destroyed Slaughtered Vaccinated  
measures

\*\*\* \*/Te

**B105 Bovine tuberculosis** Occurrence : 07/1989

Outbreaks Cases Deaths Control Destroyed Slaughtered Vaccinated  
bov measures \*/Te/qf  
Note : 225865 cattle were tested with negative results

**B106 Bovine cysticercosis** Occurrence : +?

Outbreaks Cases Deaths Control Destroyed Slaughtered Vaccinated  
bov measures \*/Su  
Note : out of 109495 cattle slaughtered 11 found infected

**B107 Dermatophilosis** Occurrence : 0000

Outbreaks Cases Deaths Control Destroyed Slaughtered Vaccinated  
\*\*\* measures \*

**B108 Enzootic bovine leukosis** Occurrence : +

Outbreaks Cases Deaths Control Destroyed Slaughtered Vaccinated  
bov 380 678 Sp/\*/  
5 Te/Su/  
Qf/Qf 704  
Note : 231468 blood samples were tested and 678 were positive

**B109 Haemorrhagic  
septicaemia** Occurrence : 10/2000

Outbreaks Cases Deaths Control Destroyed Slaughtered Vaccinated  
bov measures \*

**B110 Infectious  
bov.rhinotracheitis (IBR/IPV) Occurrence : +**

Outbreaks Cases Deaths Control Destroyed Slaughtered Vaccinated  
measures  
bov 2 9 0 Sp / \* /  
Te / qf / 8 6335  
Qf / V  
Note : 6983 blood samples were tested and 317 were positive

**B111 Theileriosis Occurrence : 0000**

Outbreaks Cases Deaths Control Destroyed Slaughtered Vaccinated  
measures  
\*\*\*\* \*

**B112 Trichomonosis Occurrence : 0000**

Outbreaks Cases Deaths Control Destroyed Slaughtered Vaccinated  
measures  
bov \* / Te  
Note : only bulls were tested in Artificial Insemination Centres

**B113 Trypanosomosis (tsetse-  
borne) Occurrence : 0000**

**B114 Malignant catarrhal  
fever Occurrence : 12/2000**

Outbreaks Cases Deaths Control Destroyed Slaughtered Vaccinated  
measures  
bov \*

**B115 Bovine spongiform  
encephalopathy Occurrence : 0000**

Outbreaks Cases Deaths Control Destroyed Slaughtered Vaccinated  
measures  
bov \* / Su /  
Qf / M

Note : 6130 cattle were tested with negative results

**B151 Ovine epididymitis  
(Brucella ovis) Occurrence : 07/1989**

Outbreaks Cases Deaths Control Destroyed Slaughtered Vaccinated  
measures  
ovi \* / Te  
Note : 3124 blood samples were tested with negative results

**B152 Cap/ovi brucel.  
(excluding B. ovis) Occurrence : 0000**

Outbreaks Cases Deaths Control Destroyed Slaughtered Vaccinated  
measures  
cap \* / Te / qf  
Note : 678 blood samples were tested with negative results  
ovi \* / Te / qf  
Note : 3518 blood samples were tested with negative results

**B153 Caprine  
arthritis/encephalitis Occurrence : +?**

Outbreaks Cases Deaths Control Destroyed Slaughtered Vaccinated  
measures  
cap \* / Te /  
Su / Qf / Qf  
Note : 199 blood samples were tested and 31 were positive , but there  
were no clinical signs of disease

**B154 Contagious agalactia Occurrence : 0000**

Outbreaks Cases Deaths Control Destroyed Slaughtered Vaccinated  
measures  
o/c \*

**B155 Contagious caprine  
pleuropneumonia Occurrence : 0000**

Control

Outbreaks Cases Deaths measures Destroyed Slaughtered Vaccinated  
cap \*

**B156 Enzootic abortion (ovine chlamydiosis) Occurrence : 0000**

Outbreaks Cases Deaths Control Destroyed Slaughtered Vaccinated  
measures \* / Te  
o/c

Note : 211 blood samples were tested with negative results

**B157 Ovine pulmonary adenomatosis Occurrence : 0000**

Outbreaks Cases Deaths Control Destroyed Slaughtered Vaccinated  
measures \*  
o/c

**B158 Nairobi sheep disease Occurrence : 0000**

Outbreaks Cases Deaths Control Destroyed Slaughtered Vaccinated  
measures \*  
ovi

**B159 Salmonellosis (S. abortusovis) Occurrence : 0000**

Outbreaks Cases Deaths Control Destroyed Slaughtered Vaccinated  
measures \*  
ovi

**B160 Scrapie Occurrence : 0000**

Outbreaks Cases Deaths Control Destroyed Slaughtered Vaccinated  
measures \* / Su /  
o/c Qf / M

Note : 4 samples were tested with negative results

**B161 Maedi-visna Occurrence : +?**

Outbreaks Cases Deaths Control Destroyed Slaughtered Vaccinated  
measures \* / Te /  
ovi Qf / Qi

Note : 2249 blood samples were tested and 308 were positive, but there were no clinical signs of disease

**B201 Contagious equine metritis Occurrence : 0000**

Outbreaks Cases Deaths Control Destroyed Slaughtered Vaccinated  
measures \*  
equ

**B202 Dourine Occurrence : 0000**

Outbreaks Cases Deaths Control Destroyed Slaughtered Vaccinated  
measures \* / Te  
equ

Note : 3203 blood samples were tested with negative results

**B203 Epizootic lymphangitis Occurrence : 0000**

Outbreaks Cases Deaths Control Destroyed Slaughtered Vaccinated  
measures \*  
equ

**B204 Encephalomyelitis (Eastern and Western) Occurrence : 0000**

Outbreaks Cases Deaths Control Destroyed Slaughtered Vaccinated  
measures \*  
equ

**B205 Equine infectious anaemia Occurrence : 10/2002**

Control

Outbreaks Cases Deaths measures Destroyed Slaughtered Vaccinated  
equ \* / Te / Qf

Note : 3566 blood samples were tested with negative results

**B206 Equine influenza** Occurrence : 0000

Outbreaks Cases Deaths Control Destroyed Slaughtered Vaccinated  
measures \* / V  
equ 925

**B207 Equine piroplasmosis** Occurrence : 0000

Outbreaks Cases Deaths Control Destroyed Slaughtered Vaccinated  
measures \*  
equ

**B208 Equine rhinopneumonitis** Occurrence : 11/2002

Outbreaks Cases Deaths Control Destroyed Slaughtered Vaccinated  
measures \* / Te / Qf / Ql / V  
equ 641

**B209 Glanders** Occurrence : 06/1996

Outbreaks Cases Deaths Control Destroyed Slaughtered Vaccinated  
measures \* / Te / Qf  
equ  
Note : 3560 blood samples were tested with negative results

**B210 Horse pox** Occurrence : 0000

Outbreaks Cases Deaths Control Destroyed Slaughtered Vaccinated  
measures \*  
equ

**B211 Equine viral arteritis** Occurrence : +?

Outbreaks Cases Deaths Control Destroyed Slaughtered Vaccinated  
measures \* / Te / Qf / Ql  
equ

Note : 234 blood samples were tested and 43 were positive, but there were no clinical signs of disease

**B212 Japanese encephalitis** Occurrence : 0000

Outbreaks Cases Deaths Control Destroyed Slaughtered Vaccinated  
measures \*  
equ

**B213 Horse mange** Occurrence : 0000

Outbreaks Cases Deaths Control Destroyed Slaughtered Vaccinated  
measures \*  
equ

**B215 Surra (Trypanosoma evansi)** Occurrence : 0000

Outbreaks Cases Deaths Control Destroyed Slaughtered Vaccinated  
measures \*  
equ

**B216 Venezuelan equine encephalomyelitis** Occurrence : 0000

Outbreaks Cases Deaths Control Destroyed Slaughtered Vaccinated  
measures \*  
equ

**B251 Atrophic rhinitis of swine** Occurrence : +?

Outbreaks Cases Deaths Control Destroyed Slaughtered Vaccinated  
measures \* / Te / Su / Qf / Ql / V  
sui 15093



avi measures \*

**B307 Fowl pox** Occurrence : 0000

Outbreaks Cases Deaths Control Destroyed Slaughtered Vaccinated  
measures \*  
avi

**B308 Fowl typhoid** Occurrence : 04/1995

Outbreaks Cases Deaths Control Destroyed Slaughtered Vaccinated  
measures \* / Te / Qf  
avi  
Note : 72425 blood samples where tested with negative results

**B309 Infect. bursal disease (Gumboro disease)** Occurrence : 12/1997

Outbreaks Cases Deaths Control Destroyed Slaughtered Vaccinated  
measures \* / Su / V  
avi  
Note : 256 blood samples were tested and 256 were positive after vaccination  
8562312

**B310 Marek's disease** Occurrence : +?

Outbreaks Cases Deaths Control Destroyed Slaughtered Vaccinated  
measures \* / Su / Qf / Ql / V  
avi  
Note : From dead tested birds 60 were positive  
5059703

**B311 Mycoplasmosis (M. gallisepticum)** Occurrence : +?

Outbreaks Cases Deaths Control Destroyed Slaughtered Vaccinated  
measures \* / Te  
avi  
Note : 386 blood samples were tested and 29 were positive

**B312 Avian chlamydiosis** Occurrence : 0000

Outbreaks Cases Deaths Control Destroyed Slaughtered Vaccinated  
measures \*  
avi

**B313 Pullorum disease** Occurrence : 04/1994

Outbreaks Cases Deaths Control Destroyed Slaughtered Vaccinated  
measures \* / Te / Qf  
avi  
Note : 72425 blood samples were tested with negative results

**B351 Myxomatosis** Occurrence : 11/1998

Outbreaks Cases Deaths Control Destroyed Slaughtered Vaccinated  
measures \* / V  
lep  
26129

**B352 Tularemia** Occurrence : 0000

Outbreaks Cases Deaths Control Destroyed Slaughtered Vaccinated  
measures \*  
lep

**B353 Rabbit haemorrhagic disease** Occurrence : 10/2000

Outbreaks Cases Deaths Control Destroyed Slaughtered Vaccinated  
measures \* / V  
lep  
21804

**B401 Viral haemorrhagic septicemia** Occurrence : 1982

Outbreaks Cases Deaths Control Destroyed Slaughtered Vaccinated  
measures \* / Qf  
pis

**B404 Spring viraemia of carp Occurrence : 0000**

Outbreaks Cases Deaths Control Destroyed Slaughtered Vaccinated  
measures \* / qf  
pis

**B405 Infectious haematopoietic necrosis Occurrence : 0000**

Outbreaks Cases Deaths Control Destroyed Slaughtered Vaccinated  
measures \* / qf  
pis

**B413 Epizootic haematopoietic necrosis Occurrence : 0000**

**B415 Oncorhynchus masou virus disease Occurrence : 0000**

**B431 Bonamiosis (*Bonamia exitiosus*, *B. ostreae*, *Mikrocytos roughleyi*) Occurrence : 0000**

**B432 MSX disease (*Haplosporidium nelsoni*) Occurrence : 0000**

**B433 Perkinsosis (*Perkinsus marinus*, *P. olseni/atlanticus*) Occurrence : 0000**

**B434 Martelliosis (*Marteilia***

***refringens*, *M. sydneyi*) Occurrence : 0000**

**B436 Mikrocytos (*Mikrocytos mackini*) Occurrence : 0000**

**B445 Taura syndrome Occurrence : -**

**B446 White spot disease Occurrence : -**

**B447 Yellowhead disease Occurrence : -**

**B451 Acariosis of bees Occurrence : 05/2000**

Outbreaks Cases Deaths Control Destroyed Slaughtered Vaccinated  
measures \* / Te / qf  
apl  
Note : 435 samples were tested with negative results.

**B452 American foulbrood Occurrence : 09/2001**

Outbreaks Cases Deaths Control Destroyed Slaughtered Vaccinated  
measures \* / Te / qf  
apl

**B453 European foulbrood Occurrence : 08/2002**

Outbreaks Cases Deaths Control Destroyed Slaughtered Vaccinated  
measures \* / Te / qf  
apl

**B454 Nosemosis of bees Occurrence : +?**

Control

Outbreaks Cases Deaths measures Destroyed Slaughtered Vaccinated

api \* / Te / Qf / Qi

Note : 435 samples were tested and 109 were positive

### B455 Varroosis

Occurrence : +

Outbreaks Cases Deaths Control Destroyed Slaughtered Vaccinated

measures

api	9	579	579	Sp / * / Te / Su / Qf / Qi	50
-----	---	-----	-----	----------------------------------	----

Note : 425 samples were tested and 40 were positive

### B501 Leishmaniosis

Occurrence : 0000

## ANIMAL HEALTH AND CONTROL MEASURES LIST C DISEASES

### C611 Listeriosis

Occurrence : 04 / 2002

Outbreaks Cases Deaths Control Destroyed Slaughtered Vaccinated

measures

bov \* / Te / Qi  
Note : 1256 cattle blood samples were tested and 1 were positive

\* / Te

cap Note : 160 goats blood samples were tested with negative results

\* / Te

ovi Note : 1982 sheep blood samples were tested and 2 were positive

### C612 Toxoplasmosis

Occurrence : 0000

Outbreaks Cases Deaths Control Destroyed Slaughtered Vaccinated

measures

bov \*  
sui \*

### C613 Melioidosis

Occurrence : 0000

Outbreaks Cases Deaths Control Destroyed Slaughtered Vaccinated

measures

ovi \*

### C614 Blackleg

Occurrence : 0000

Outbreaks Cases Deaths Control Destroyed Slaughtered Vaccinated

measures

bov \*

### C615 Botulism

Occurrence : -

Outbreaks Cases Deaths Control Destroyed Slaughtered Vaccinated

measures

\*\*\* \*

### C616 Other clostridial infections

Occurrence : 0000

Outbreaks Cases Deaths Control Destroyed Slaughtered Vaccinated

measures

\*\*\* \*

### C617 Other pasteurellosis

Occurrence : 11 / 2000

Outbreaks Cases Deaths Control Destroyed Slaughtered Vaccinated

measures

bov \*

sui \* / V

1289

### C618 Actinomycosis

Occurrence : 0000

Outbreaks Cases Deaths Control Destroyed Slaughtered Vaccinated

measures

bov \*

**C619 Intestinal Salmonella infections** Occurrence : 08/2002

Outbreaks Cases Deaths Control Destroyed Slaughtered Vaccinated  
measures \*  
bov \*  
sui \* / V 479

**C620 Coccidiosis** Occurrence : +?

Outbreaks Cases Deaths Control Destroyed Slaughtered Vaccinated  
measures \*  
avi \*

**C621 Distomatosis (liver fluke)** Occurrence : 0000

Outbreaks Cases Deaths Control Destroyed Slaughtered Vaccinated  
measures \*  
\*\*\*

**C622 Filariosis** Occurrence : 0000

Outbreaks Cases Deaths Control Destroyed Slaughtered Vaccinated  
measures \*  
\*\*\*

**C652 Mucosal disease/Bovine virus diarrhoea** Occurrence : +

Outbreaks Cases Deaths Control Destroyed Slaughtered Vaccinated  
measures  
bov 1 2 0 Sp / \* /  
Te / Qf / Qi 2  
Note : 6965 blood samples were tested and 789 were positive(antibody)

**C653 Vibriotic dysentery** Occurrence : 11/2001

Outbreaks Cases Deaths Control Destroyed Slaughtered Vaccinated  
measures

sui \*

**C654 Warble infestation** Occurrence : -

Outbreaks Cases Deaths Control Destroyed Slaughtered Vaccinated  
measures \*  
bov \*

**C701 Contagious pustular dermatitis** Occurrence : 0000

Outbreaks Cases Deaths Control Destroyed Slaughtered Vaccinated  
measures \*  
\*\*\*

**C702 Foot-rot** Occurrence : 0000

Outbreaks Cases Deaths Control Destroyed Slaughtered Vaccinated  
measures \*  
\*\*\*

**C703 Contagious ophthalmia** Occurrence : 0000

Outbreaks Cases Deaths Control Destroyed Slaughtered Vaccinated  
measures \*  
ovi \*

**C704 Enterotoxaemia** Occurrence : 0000

Outbreaks Cases Deaths Control Destroyed Slaughtered Vaccinated  
measures \*  
\*\*\*

**C705 Caseous lymphadenitis** Occurrence : 0000

Outbreaks Cases Deaths Control Destroyed Slaughtered Vaccinated  
measures \*  
\*\*\*

**C706 Sheep mange** **Occurrence : 0000**  
 Outbreaks Cases Deaths Control Destroyed Slaughtered Vaccinated  
 ovi measures \* \*

**C751 Equine coital exanthema** **Occurrence : 0000**  
 Outbreaks Cases Deaths Control Destroyed Slaughtered Vaccinated  
 equ measures \* \*

**C752 Ulcerative lymphangitis** **Occurrence : 1947**  
 Outbreaks Cases Deaths Control Destroyed Slaughtered Vaccinated  
 equ measures \* \*

**C753 Strangles** **Occurrence : 12/2000**  
 Outbreaks Cases Deaths Control Destroyed Slaughtered Vaccinated  
 equ measures \*

**C754 Salmonellosis (S. abortusequi)** **Occurrence : 0000**  
 Outbreaks Cases Deaths Control Destroyed Slaughtered Vaccinated  
 equ measures \*

**C801 Swine erysipelas** **Occurrence : 03/1992**  
 Outbreaks Cases Deaths Control Destroyed Slaughtered Vaccinated  
 sui measures \* / v 180040

**C851 Infectious coryza** **Occurrence : 0000**  
 Outbreaks Cases Deaths Control Destroyed Slaughtered Vaccinated  
 avi measures \* \*

**C853 Avian encephalomyelitis** **Occurrence : 0000**  
 Outbreaks Cases Deaths Control Destroyed Slaughtered Vaccinated  
 avi measures \* \*

**C854 Avian spirochaetosis** **Occurrence : 0000**  
 Outbreaks Cases Deaths Control Destroyed Slaughtered Vaccinated  
 avi measures \*

**C855 Avian salmonellosis** **Occurrence : +**  
 Outbreaks Cases Deaths Control Destroyed Slaughtered Vaccinated  
 avi 3 85 23 S / \* / Su / Qf / Qi 62

**C856 Avian leukosis** **Occurrence : -**  
 Outbreaks Cases Deaths Control Destroyed Slaughtered Vaccinated  
 avi measures \*

**CASES OF ANIMAL DISEASES IN HUMAN POPULATION**

CODE OF DISEASE	NAME OF DISEASE	NUMBER OF HUMAN CASES	NOTE
A080	Rift Valley fever	0	
B051	Anthrax	0	
B053	Echinococcosis/hydatidosis	0	
B056	Leptospirosis	10	

B057	Q fever	0
B058	Rabies	1
B060	New world screwworm ( <i>C. hominivorax</i> )	0
B061	Old world screwworm ( <i>C. bezziana</i> )	0
B062	Trichinellosis	0
B105	Bovine tuberculosis	22
B106	Bovine cysticercosis	0
B209	Glanders	0
B212	Japanese encephalitis	0
B216	Venezuelan equine encephalomyelitis	0
B252	Porcine cysticercosis	0
B312	Avian chlamydiosis	0
B352	Tularaemia	0
B501	Leishmaniasis	0
C611	Listeriosis	0
C612	Toxoplasmosis	8
C801	Swine erysipelas	3
E001	Brucellosis	0
E002	Salmonellosis	0
		799

**Note :**

**ANNUAL WRITTEN REPORT**

**I. New activities of the veterinary services**

There are some significant changes have been made in the structure of Food and Veterinary service in autumn 2003.

Animal health and welfare department was renamed to Veterinary surveillance department.

Animal health surveillance division were renamed to Animal infectious diseases surveillance division under which two new Epizootic control sector and Zoonoses control sector.

As a becoming member state of European Union (EU), Latvia was continuing to transpose EU legislation with regard to infectious diseases

**II. Comments on specific List A diseases**

Latvia remained free from list A diseases  
Diseases never reported:  
Vesicular Stomatitis

- Swine vesicular disease
- Peste des petits ruminants
- Lymphy skin disease
- Rift valley fever
- Biloungue
- Sheep pox and goat pox
- African horse sickness
- African swine fever
- Highly pathogenic avian Influenza
- Newcastle disease

Diseases reported absent in 2003 (in parentheses: date of last outbreak):

- Foot and mouse disease 11/1987
- Classical swine fever 04/1996
- Contagious bovine pleuropneumonia 1922
- Rinderpest 1921

The monitoring of List A diseases (excluding Lymphy skin disease and Rift Valley fever) was provided in all territory of Latvia during 2003.

All results were negative.

The specialists of the FVS were prepared the informative bulletins concerning Classical swine fever and Foot and Mouse disease for animal owners and keepers.

**III. Comments on specific List B diseases**

Diseases reported absent in 2003 (in parentheses: date of last outbreak):

- Bovine tuberculosis: (07/1989)
- Swine tuberculosis (08/1984)
- Avian tuberculosis (01/1993)
- Bovine brucellosis (1963)
- Swine brucellosis (12/1994)
- Sheep and goat brucellosis never reported
- Ovine epididymitis (07/1989)

Multiple species diseases

**1. Rabies**

A total 964 cases of rabies were registered in all 26 districts. The main carriers and keepers of Rabies in the nature are wild predators (foxes, raccoon-dogs, badgers and other wild animals). Foxes and raccoon-dogs play central role in spreading of Rabies in Latvia. 471 foxes (48,9%) 285 raccoon-dogs (29,6%) and 33 badgers (3,4%) were found sick with Rabies from all registered sick animals during 2003. Every year

the epizootic process of Rabies involves domestic animals and humans.

One human case was registered in 2003.

Among domestic animals the following number and species were included: 62 dogs (6,4%), 52 cats (5,4%) and 21 cattle (2,2%).

Cats and dogs in Latvia are obligated to prophylactic vaccination against Rabies once per year. In cases when domestic animals have been in contact with wild animals suspected on Rabies or infected, the obligatory vaccination against Rabies has been carried out for all animals in the farm. During 2003 oral vaccination of foxes and raccoon-dogs was carried out in all 26 administrative districts.

There were two vaccination campaigns (summer and autumn) organized and 300 000 vaccine baits were distributed in the territory of Latvia, but due to financial reasons, baits were not distributed as many as it would be necessary having regard our fox population.

The specialists of the FVS were issued the Informative bulletins concerning Rabies for animal owners.

#### Cattle diseases

##### 1. Bovine spongiform encephalopathy

As a becoming member state of European Union (EU), Latvia has transposed EU legislation with regard to BSE.

The system of BSE surveillance is improved during 2003. The total number of sampled animal in 2003 was 6126 from which 1014 were fallen stock, 263 emergency slaughtered animals and 4838 healthy slaughtered bovine animals. 11 suspects to BSE were registered in 2003. 54 tested animals were imported from countries with indigenous BSE cases. All BSE testing results were negative. There is no any positive case of BSE found so far.

The specialists of the FVS issued the informative bulletins concerning BSE.

The training courses were organized for inspectors of territorial FVS offices to improve theoretical and practical skills concerning BSE clinical signs and sampling procedures.

##### 2. Enzootic bovine leucosis, bovine brucellosis,

According to  Animal infectious diseases state surveillance programme 2003  to enzootic bovine leucosis (EBL) serological testing were subjected:

" certified breeding bulls twice per year;

" heifers from 12 months of age (in EBL free herds) once per year;

" cows and breeding bulls (in EBL free herds) once per year blood sampling or testing of bulk milk sample twice per year;

" bovine animals during 30 days before placing in a new herd;

From 231 468 serologically tested bovine animals 678 were positive to leucosis (0,29%). In three administrative districts disease has not been registered.

Cattle in affected herds are subjected to serological testing three times per year. All positive animals were slaughtered. As a result of disease eradication programme we hope to reach disease free status in a very near future.

FVS has started to test bulk milk samples to enzootic bovine leucosis and bovine brucellosis in 2002.  
208 135 bovine animals were tested negative to brucellosis during 2003.

#### Swine diseases

##### 1. Aujeszky disease

9143 animals were serologically tested and 435 were found positive to Aujeszky disease, but clinical signs were not detected.

80304 animals were vaccinated against Aujeszky disease with marked vaccine.

Latvia has no state surveillance programme for Aujeszky disease. Eradication of disease is carried out on voluntary basis. Vaccination of animals requires large finance funds, but not all the farmers can presume it.

#### IV. Comments on other diseases

Approved by  
Food and Veterinary Service  
of the Republic of Latvia  
Order No.234  
May 24, 2002

Issued on the basis of  
the Law on Veterinary Medicine  
Paragraph 4, Article 3  
and Paragraph 16, sub paragraph 2

## **INSTRUCTION ON NOTIFICATION OF HIGHLY DANGEROUS ANIMAL INFECTUOUS DISEASES**

### **I General items**

1. The instruction determines the order of collection and notification of information in the Republic of Latvia on the following issues:

- 1.1. suspicion on outbreak of highly dangerous animal infectious disease;
- 1.2. confirmation of the outbreak of highly dangerous animal infectious disease;
- 1.3. results of laboratory examination concerning highly dangerous animal infectious disease;
- 1.4. eradication measures of highly dangerous animal infectious disease;
- 1.5. cancellation of restrictions applied concerning outbreak of highly dangerous animal infectious disease.

2. The Central Authority of Food and Veterinary Service (further in the text – FVS) in accordance with established procedure shall receive the information mentioned in Article 1 of the present instruction from:

- 2.1. FVS territorial departments;
- 2.2. State Veterinary Medicine Diagnostics Centre.

3. Territorial departments of FVS shall receive the information on suspicion or outbreak of highly dangerous animal infectious disease from:

- 3.1. state authorised veterinarians;
- 3.2. private veterinary practitioners.

4. The Central Authority of FVS in accordance with established procedure shall notify the information mentioned in Article 1 of the present instruction to:

- 4.1. Office International des Epizooties (OIE);
- 4.2. European Commission<sup>1</sup> and Member States of European Union (EC)<sup>1</sup>;
- 4.3. World Health Organization (WHO);
- 4.4. United Nations Food and Agriculture Organization (FAO);
- 4.5. countries with which Latvia has land border;
- 4.6. countries having agreement with Latvia about notification of outbreak of highly dangerous animal infectious diseases and cancellation of restrictions applied concerning outbreak of highly dangerous animal infectious diseases.

### **II Order of operative notification**

**of highly dangerous animal infectious diseases within the Republic of Latvia**

5. State authorised veterinarian or private veterinary practitioner establishing contamination of one or more animals of infectious diseases mentioned in Annex 1 of the present instruction, shall immediately by telephone inform a territorial department of FVS and give instructions to animal owner about restrictions concerning potential outbreak of highly dangerous animal infectious disease.

6. Territorial department of FVS shall record the received notification in the "Journal of registration of suspicion of highly dangerous animal infectious disease". Journal shall be set up in accordance with Annex 9 of the present instruction.

7. Territorial department of FVS upon receipt notification mentioned in Article 5 of the present instruction about suspicion of contamination of animals with any of disease mentioned in Annex 1 of the present instruction, shall send completed form mentioned in Annex 2 of the present instruction by facsimile to the Central authority of FVS and State Veterinary Medicine Diagnostics Centre, as well as immediately inform by telephone:

- 7.1. the territorial department of FVS;
- 7.2. the State Veterinary Medicine Diagnostic Centre;
- 7.3. the communication service of the State Fire Fighting and Rescue Service city/regional department;
- 7.4. the branch of the territorial Public Health Agency in accordance with order provided by regulatory documents.

8. By sending samples for laboratory examination shall complete form for sending of samples for laboratory examination in accordance with Annex 3 of the present instruction. Annex 3 shall not apply to cases when sending samples for examinations on Scrapie and Bovine spongiform encephalopathy.

9. Territorial department of FVS upon receipt of notification on information mentioned in Article 4 of the present instruction, in accordance with the requirements provided by regulatory documents shall prepare notification for animal owners about temporary restrictions applied to animal holding (see Annex 4 of the present instruction).

10. Territorial department of FVS shall prepare the operative notification on suspicion of outbreak of highly dangerous animal infectious diseases (see Annex 5 of the present instruction) and send a copy by facsimile, e-mail or by other means to the Central authority of FVS.

11. In case of confirmed laboratory diagnose on any of the disease mentioned in Annex 1 of the present instruction State Veterinary Medicine Diagnostics Centre shall immediately by telephone and facsimile, e-mail or by other means sending the results of laboratory investigations shall notify to:

- 11.1. the territorial department of FVS where sample has been taken;
- 11.2. the Central authority of FVS.

12. Territorial department of FVS upon receipt of positive results of laboratory investigation of any of the disease mentioned in Annex 1 of the present instruction and by imposing restrictions to the affected farm where disease has been confirmed shall follow the form given in Annex 4 of the present instruction.

13. Territorial department of FVS shall collect all the information and fill in the form of operative notification (Annex 6) and shall immediately send it by facsimile, e-mail or by other means to:

13.1. the Central authority of FVS;

13.2. the communication service of the State Fire Fighting and Rescue Service city/regional department;

13.3. the branch of territorial Public Health Agency in accordance with order provided by regulatory documents.

14. Territorial department of FVS shall once a week inform the Central authority of FVS about all the measures applied and carried out in case of outbreak of disease mentioned in Annex 1 of the present instruction as well as shall notify about results of epizootic (epidemiological) investigation until all the restrictions applied in case of outbreak of highly dangerous infectious disease are cancelled.

15. Territorial department of FVS upon receipt of negative results of laboratory investigation of any of the disease mentioned in Annex 1 of the present instruction by cancelling the temporary restrictions to holding with suspected animals shall complete the form in accordance with Annex 7 of the present instruction and send it by facsimile, e-mail or by other means to:

15.1. the Central authority of FVS;

15.2. the communication service of the State Fire Fighting and Rescue Service city/regional department;

15.3. the branch of territorial Public Health Agency in accordance with order provided in regulatory documents.

16. Territorial department of FVS upon cancellation of restrictions applied to farm in case of outbreak of highly dangerous animal infectious disease shall complete form in accordance with Annex 8 of the present instruction and send it by facsimile, e-mail or by other way to:

16.1. the Central authority of FVS;

16.2. the communication service of the State Fire Fighting and Rescue Service city/regional department;

16.3. if necessary – the branch of territorial Public Health Agency.

**Highly dangerous animal infectious diseases**

<b>Code Nr.</b>	<b>Name of the disease</b>
A010	<b>Foot and mouth disease (FMD)</b>
A011	FMD – Virus O
A012	FMD – Virus A
A013	FMD – Virus C
A014	FMD – Virus SAT1
A015	FMD – Virus SAT2
A016	FMD – Virus SAT3
A017	FMD – Virus Asia1
A018	FMD – Virus not typed
A020	Vesicular Stomatitis (VS)
A021	VS – Virus Indiana
A022	VS – Virus New Jersey
A023	VS – Virus not typed
A030	<b>Swine Vesicular Disease</b>
A040	Rinderpest
A050	<b>Peste des Petits Ruminats</b>
A060	Contagious Bovine Pleuropneumonia
A070	<b>Lumpy Skin Disease</b>
A080	<b>Rift Valley Fever</b>
A090	<b>Bluetongue</b>
A100	<b>Sheep and Goat Pox</b>
A110	African Horse Sickness
A120	<b>African swine fever</b>
A130	Classical swine fever (Hog cholera)
A150	<b>Fowl plaque</b>
A160	Newcastle disease
B051	Anthrax
B115	Bovine spongiform encephalopathy
<b>B160</b>	Scrapie
B256	Enterovirus encephalomyelitis

## Notification on suspicion of contamination of animals

Name of the territorial department of FVS \_\_\_\_\_

\_\_\_\_\_  
**Name, surname of person who has received the notification** \_\_\_\_\_

\_\_\_\_\_  
**Name, surname and telephone number of the person who has notified about the suspected contamination of animals** \_\_\_\_\_

\_\_\_\_\_  
**Date and time when the notification was received** \_\_\_\_\_

**Information about the suspected holding** (address, telephone number, herd registration number, name, surname and address of the animal / shed owner):

\_\_\_\_\_  
\_\_\_\_\_

\_\_\_\_\_  
**Name, surname, address, telephone number, license number of veterinarian**

\_\_\_\_\_

\_\_\_\_\_  
**Species, categories and number of animals within the holding**

\_\_\_\_\_

\_\_\_\_\_  
**Anamnesis, clinical signs, results of postmortem examination, number and age of the sick and dead animals** \_\_\_\_\_

\_\_\_\_\_

\_\_\_\_\_  
**Registration number of the suspected case** \_\_\_\_\_

**Name, surname of State Senior Veterinary Inspector / State Veterinary Inspector, (delete as appropriate), who has been notified about the suspicion**

\_\_\_\_\_

\_\_\_\_\_  
**Date and time when notification has been reported to State Senior Veterinary Inspector / State Veterinary Inspector, (delete as appropriate)**

\_\_\_\_\_  
**The evaluated time when State Senior Veterinary Inspector / State Veterinary Inspector, (delete as appropriate) shall give the first report about the situation in the suspected farm** \_\_\_\_\_

**Signature** \_\_\_\_\_ **Date, time** \_\_\_\_\_

**Form for sending of samples for laboratory investigation No. \_\_\_\_\_**

Name of the territorial department of FVS \_\_\_\_\_

1. Registration number of the suspected case \_\_\_\_\_

2. Address of the holding \_\_\_\_\_

3. Telephone No. \_\_\_\_\_

4. Registration number of the herd \_\_\_\_\_

5. Anamnesis:

5.1. date when the notification on suspicion was received \_\_\_\_\_

5.2. date when the first clinical signs of the disease were established \_\_\_\_\_

5.3. clinical signs \_\_\_\_\_

5.4. results of post mortem examination \_\_\_\_\_

5.5. possibly source of infection \_\_\_\_\_

6. Susceptible animals in the holding

Species / category	Number of animals	Number of sick animals	Number of dead animals
Bulls			
Milking cows			
Other cows			
Young stock (cows)			
calves			
<b>Total number of cows</b>			
Sheep			
Young stock (sheep)			
<b>Total number of sheep</b>			
Goats			
Young stock (goats)			
<b>Total number of goats</b>			
Boars			
Sows			
Young stock (pigs)			
Fattening pigs			
Waned pigs			
Piglets			
<b>Total number of pigs</b>			

7. Information about the samples delivered for laboratory investigation

Pathological material	Animal (species/category)	Animal identification No.	Sample No.

8. Date and time when the sample has been sent to the laboratory \_\_\_\_\_

9. Manner of sample transportation to the laboratory \_\_\_\_\_

10. Name, surname, license No, address of the workplace and telephone number of  
veterinarian who took samples for laboratory investigation \_\_\_\_\_

\_\_\_\_\_

\_\_\_\_\_

\_\_\_\_\_

\_\_\_\_\_

\_\_\_\_\_

name, surname

\_\_\_\_\_

position

\_\_\_\_\_

signature

Notification about the temporary introduction of restrictions

Name of the territorial department of FVS \_\_\_\_\_  
\_\_\_\_\_

Registration number of the suspected case \_\_\_\_\_

To (owner of the holding / animal holder) \_\_\_\_\_

Address \_\_\_\_\_  
\_\_\_\_\_

Reason of issuing the notification \_\_\_\_\_

“I, the undersigned \_\_\_\_\_

(name of the territorial department of FVS)

State Senior Veterinary Inspector / State Veterinary Inspector (delete as appropriate) hereby inform you, that in accordance with the requirements provided by regulatory documents in the case of suspected outbreak of \_\_\_\_\_ until receiving of the results of

(name of the disease)

laboratory investigation, all the persons, animals, products of animal origin, materials and means of transport in the holding or outside the holding shall be subjects, which are subjected to the requirements enclosed in Annex of the present notification\*”.

Name, surname of inspector \_\_\_\_\_

Signature \_\_\_\_\_ date, time \_\_\_\_\_

**\* Shall mention all requirements / restrictions as Annex of the present notification.**

**Operative notification “On suspicion of the outbreak of highly dangerous animal infectious disease”**

Name of the territorial department of FVS: \_\_\_\_\_

The responsible veterinarian: \_\_\_\_\_

**Registration No. of the suspected case:** \_\_\_\_\_

**Date:** \_\_\_\_\_

I Animal owner

Name, surname: \_\_\_\_\_

**Address:** \_\_\_\_\_

**Telephone No.:** \_\_\_\_\_

**Number and species of animals in the farm:**

<b>Bulls</b>	<b>Cows</b>	<b>Heifers</b>
<b>Fattening cattle</b>	<b>Calves, younger than 6 months</b>	
<b>Boars</b>	<b>Sows</b>	
<b>Piglets</b>	<b>Gilts</b>	<b>Fattening pigs</b>
<b>Sheep</b>	<b>Goats</b>	<b>Horses</b>
<b>Fowls</b>	<b>Other animals</b>	

**II Person who notified about the contamination of animals (name, surname):**

\_\_\_\_\_

Date: \_\_\_\_\_  
Motivation of the notified suspicion (clinical signs): \_\_\_\_\_

\_\_\_\_\_

**State senior veterinary inspector / state veterinary inspector** (delete as appropriate):  
visited the holding on: date \_\_\_\_\_ time \_\_\_\_\_

**Number of the clinically sick animals:** \_\_\_\_\_

\_\_\_\_\_

**Number of the possibly contaminated animals at pasture:** \_\_\_\_\_  
**Clinical symptoms:** \_\_\_\_\_

\_\_\_\_\_

**Primary diagnose:** \_\_\_\_\_

\_\_\_\_\_

**Material sent for laboratorial investigations:** \_\_\_\_\_

\_\_\_\_\_

**Results of laboratorial investigations:** \_\_\_\_\_

\_\_\_\_\_

**Information received from the animal owner about the contamination of animals**

**Time when the first symptoms were observed:** \_\_\_\_\_

**Clinical signs:** \_\_\_\_\_

\_\_\_\_\_

**Location of the sick animals at the time when signs of contamination were observed :**

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**III Location of the holding:** \_\_\_\_\_

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**NB! Location of the sick animals in the farm has to be schematically indicated in the last page of the current notification!**

**The closest farm breeding agricultural animals (name, surname of the owner, place of location):** \_\_\_\_\_

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**All agricultural animal holdings within the radius of 500 meters around the possibly affected place :** \_\_\_\_\_

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**Manure – description of methods of storage, by indicating their possible contact with water sources:** \_\_\_\_\_

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**Sewerage system (description, and whether there is a possibility of sewage to come into any of water reservoirs):** \_\_\_\_\_

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**Distance to the forest:** \_\_\_\_\_  
**Indicate the distance to the closest populated places, summer cottages, allotments, camp sites (if any):** \_\_\_\_\_

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**Roads (and paths) leading to / from / along the possibly affected farm:** \_\_\_\_\_

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#### IV Feeding / Watering

Water supply: \_\_\_\_\_

Indicate the feedingstuffs used to feed the animals during the last 30 (thirty) days until the day of emergence of clinical signs of the disease: \_\_\_\_\_

The place where animals are fed and place where feedingstuffs are stored: \_\_\_\_\_

#### V The possible contacts within 21 (twenty-one) days before emergence of the first clinical signs of the disease:

##### A. Purchase of animals:

- species of animals, number of animals and date when the animals were purchased: \_\_\_\_\_

veterinary certificate (No., the issuing authority) / declaration of animal movement: \_\_\_\_\_

the farm of origin: \_\_\_\_\_

- name, surname, address and telephone No. of the mediator / seller (if the animal has been bought from the mediator): \_\_\_\_\_

- whether the sick animals are among the purchased animals: \_\_\_\_\_

##### B. Selling of animals:

##### 1) Animals sold to the other herd:

- species, number, date: \_\_\_\_\_

veterinary certificate (No., the issuing authority) / declaration of animal movement: \_\_\_\_\_

name, surname and address of the receiver of animals: \_\_\_\_\_

- name, surname, address and telephone No. of the mediator / seller (if the animal has been bought from the mediator): \_\_\_\_\_  
\_\_\_\_\_  
\_\_\_\_\_

2) animals sold for slaughtering:

- species, number, date: \_\_\_\_\_  
\_\_\_\_\_
- veterinary certificate (No., the issuing authority) / declaration of animal movement: \_\_\_\_\_  
\_\_\_\_\_
- the slaughterhouse, where animals are transported: \_\_\_\_\_  
\_\_\_\_\_
- through the market (name of the market): \_\_\_\_\_  
supplier / mediator (name, surname, address, telephone No.): \_\_\_\_\_  
\_\_\_\_\_

**NB! The parking place of the transport vehicle, used for animal transportation has to be marked in the location plan of the holding on the last page of the present notification form!**

3) animals moved out of the farm due to other reasons (animal exhibition / trade / medical treatment in the clinic, etc.):

- species, number, date: \_\_\_\_\_  
\_\_\_\_\_
- veterinary certificate (No., issuing authority) / animal movement declaration: \_\_\_\_\_  
\_\_\_\_\_
- receiver: \_\_\_\_\_
- whether the animal was returned to the holding (date): \_\_\_\_\_

**C. Products of animal origin:**

1) Realization of milk:

- frequency \_\_\_\_\_
- the last date of realization \_\_\_\_\_
- the milk from the farm is transported to (where, for whom) \_\_\_\_\_  
\_\_\_\_\_
- whether the driver has been in the farm \_\_\_\_\_
- whether the milk has been realized to a private person \_\_\_\_\_  
\_\_\_\_\_

**NB! The parking place of transport vehicle, used for the transportation of milk, has to be marked in the location plan of the farm on the last page of the present notification form!**

2) Purchase / selling of other products:

- Wool: \_\_\_\_\_
- Other products: \_\_\_\_\_

\_\_\_\_\_

D. Feedingstuffs

Date of purchase / selling and purchaser / seller: \_\_\_\_\_

\_\_\_\_\_

1) concentrated fodder and other additives: \_\_\_\_\_

\_\_\_\_\_

2) succulent fodder: \_\_\_\_\_

\_\_\_\_\_

3) hay, straw: \_\_\_\_\_

\_\_\_\_\_

E. Technical equipment (including feeding facilities etc.)

Technical equipment has been purchased / borrowed (from whom, where): \_\_\_\_\_

\_\_\_\_\_

type of technical equipment: \_\_\_\_\_

date: \_\_\_\_\_

fertilization and other, by the use of technical equipment (address, telephone No.): \_\_\_\_\_

\_\_\_\_\_

date of fertilization: \_\_\_\_\_

whether there is any technical equipment at the farm's disposal and how this equipment is being used: \_\_\_\_\_

\_\_\_\_\_

F. Persons who have visited the farm

1) veterinarian:

- date of the visit: \_\_\_\_\_

- purpose of the visit: \_\_\_\_\_

- name and surname of the veterinarian: \_\_\_\_\_

2) Artificial inseminator:

- name of artificial inseminator: \_\_\_\_\_

- date of the visit and number of the artificially inseminated animals: \_\_\_\_\_

\_\_\_\_\_

3) Other persons (consultant, auxiliary worker, technician, blacksmith, and others):

- date of the visit: \_\_\_\_\_

- purpose of the visit: \_\_\_\_\_
  - name, surname of the visitor: \_\_\_\_\_
  - whether a person has been in contact with any of the animal in the farm: \_\_\_\_\_
- \_\_\_\_\_
- \_\_\_\_\_

**G. Information regarding persons / workers living in the farm:**

- whether the animal owner / workers have visited other farms where animals are bred:
- \_\_\_\_\_
- \_\_\_\_\_
- \_\_\_\_\_

- whether any member of the family, tenants and others are working outside the farm where there is a possibility to come into direct/indirect contact with animals (other farms, slaughter houses, dairies, etc.):
- \_\_\_\_\_
- \_\_\_\_\_
- \_\_\_\_\_
- \_\_\_\_\_
- \_\_\_\_\_

- whether any of the person living or working in the farm has been in a foreign country (date of the visit): \_\_\_\_\_
- \_\_\_\_\_
- \_\_\_\_\_
- \_\_\_\_\_

**VI. Information concerning other animals:**

**A.**

- dogs, number: \_\_\_\_\_
  - cats, number: \_\_\_\_\_
  - other animals, number: \_\_\_\_\_
  - whether any of the above-mentioned animal has participated in an exhibition, competition or other similar event: \_\_\_\_\_
- \_\_\_\_\_
- \_\_\_\_\_
- \_\_\_\_\_

**B.**

- strayed dogs and cats, the approximate number: \_\_\_\_\_
  - whether they are fed: \_\_\_\_\_ where \_\_\_\_\_
- \_\_\_\_\_

- **rodents:** \_\_\_\_\_
- **has the plan of rat extermination been developed:** \_\_\_\_\_
- **wild birds:** \_\_\_\_\_
- **blood-sucker insects:** \_\_\_\_\_

**C.**

**Presence of wild animals:** \_\_\_\_\_

**The plan of the holding and its territory:**

The plan should comprise the following information: ventilation openings (including each broken or open window), points of water intake, heaps of manure, facilities for the storage of manure, facilities for the storage of liquid manure, sick animals, transport vehicles for the slaughterhouse, means of transport for the transportation of milk, roads, water elements, etc.

**The State Senior Veterinary Inspector / State Veterinary Inspector** (delete as appropriate):

\_\_\_\_\_  
(signature, name and surname)

(seal)



**15. The causing agent of the disease:**

**16. Possible source of the infection:**

**17. Type of spreading of the infection:**

**18. Number of sick animals during notification:**

**Additional information** (date when the primary contamination has been established; number of the outbreaks associated with the primary outbreak; type of the outbreak of the infectious disease (enzootic, sporadic, epizootic, panzootic); description of the region or rural municipality (describe the place of the affected territory, its size and the neighbouring regions); information about other regions subjected to restrictions):

**Control /eradication measures during the time of notification:**

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Name and surname of the State Senior Veterinary Inspector / State Veterinary Inspector  
(delete as appropriate)

---

Signature

Seal

**Cancellation of the operative notification “On suspicion of the outbreak of highly dangerous animal infectious disease”**

1. Date: \_\_\_\_\_
2. Name of the territorial department of FVS: \_\_\_\_\_
3. Address, telephone number: \_\_\_\_\_  
\_\_\_\_\_
4. Name and code of the infectious disease: \_\_\_\_\_  
\_\_\_\_\_
5. Address of the place, where restrictions shall be cancelled: \_\_\_\_\_  
\_\_\_\_\_
6. Name of the laboratory where laboratory investigations have been carried out: \_\_\_\_\_  
\_\_\_\_\_
7. Number and conclusion of the laboratory expertise: \_\_\_\_\_  
\_\_\_\_\_  
\_\_\_\_\_

\_\_\_\_\_  
Name and surname of the State Senior Veterinary Inspector / State Veterinary Inspector  
(delete as appropriate)

\_\_\_\_\_  
Signature

Seal

**Cancellation of the operative notification “On confirmation of the outbreak of highly dangerous animal infectious disease”**

1. Date: \_\_\_\_\_

2. Name of the territorial department of FVS: \_\_\_\_\_

3. Address, telephone No: \_\_\_\_\_

4. Name and code of the animal infectious disease: \_\_\_\_\_

5. Address of the place where restrictions shall be cancelled:

\_\_\_\_\_

6. Number of the destroyed animals and the date when animals have been destroyed:

\_\_\_\_\_

7. The final cleaning and disinfection of the holding has been carried out: \_\_\_\_\_

(date, the used disinfectant, concentration)

(disinfected area m<sup>2</sup>, time of exposition of the disinfectant, the amount of disinfectant used per 1 m<sup>2</sup>)

(name, surname, position of the person who carried out disinfection)

(the result of laboratory quality control of disinfection)

8. Rat extermination has been carried out on \_\_\_\_\_ (date)

by \_\_\_\_\_  
(name of the establishment or name, surname, position of the person who carried out rat extermination)

\_\_\_\_\_  
Name, surname of the State Senior Veterinary Inspector / State Veterinary Inspector  
(delete as appropriate)

\_\_\_\_\_  
Signature

\_\_\_\_\_  
Seal

Journal of registration of suspicion of highly dangerous animal infectious diseases

No.	Date and time, when the notification was received	Name of the disease	The farm / holding, (address, owner, telephone No.) where suspicion has established	The person who notified (name, surname, address, telephone No.)	The person who received the notification (name, surname, position)	Remarks*

\* For supplementary information, received from the person who notifies about suspected contamination of animals (species of the animals, number of the animals, clinical signs etc.).

Approved by  
Food and Veterinary  
Service  
May 09, 2002 Order No  
195

## **Instruction on disinfection of animal holdings, rehabilitation of products, materials and surroundings**

### **I General provisions**

Disinfection and rehabilitation comprise a set of measures including mechanical cleaning of vehicles, holdings, auxiliary premises and surroundings; washing of holdings, auxiliary premises and equipment (sanitary recondition in the holding and auxiliary premises); elimination of the pathogen using effective disinfection agents; biothermal treatment of manure; disinfection of slurry and sewage; treatment of fodder; treatment of milk, meat, skin, etc. to eliminate the pathogen.

Specific disinfection requirements with regards to defined infectious diseases are in the process of preparation.

### **II Disinfection methods**

#### **1. The following methods are used for disinfection:**

- 1.1. Chemical;
- 1.2. Physical.

#### **2. Chemical method:**

2.1. Traditional or in Latvia registered disinfection substances, which eliminate the pathogen of the given disease, are used. Principles determining the choice of the traditional disinfection substances are described in Appendix 1 to this instruction.

2.2. Chemical disinfection substances shall be used according to the user's instructions;

2.3. Specific characteristics of the pathogen, type of disinfection objects, temperature in the premises and other factors shall be taken into account upon determining concentration, volume and time of exposition of the disinfection substance.

2.4. Work security requirements (including use of specific clothing) shall be observed during disinfection;

2.5. Alkaline disinfection substances shall be used heated;

2.6. Chemical disinfection substances may harm painted surfaces and cause corrosion of metallic objects.

#### **3. Physical method:**

3.1. The pathogen is inactivated by treating metallic objects with an open flame, burning litter, fodder, dry manure, carcasses, etc.;

3.2. In products of animal origin, such as meat and milk, pathogens of various diseases are eliminated by the use of high temperature – pasteurization and sterilizations.

### III Disinfection types

#### 4. Types of disinfection:

4.1. Prophylactic disinfection;

4.2. Compulsory disinfection:

4.2.1. preliminary disinfection;

4.2.2. final disinfection.

**5. Prophylactic disinfection** – is carried out with an aim to eliminate the pathogen in time.

5.5. If there is a threat of an outbreak of an highly contagious disease specific prophylactic measures including disinfection of vehicles and footwear according to Food and Veterinary Service instructions are applied on the state border, roads, ports, etc.

5.2. Animal holdings, auxiliary premises, animal care equipment that has been in direct contact with the animals, as well as enclosures, shall be disinfected;

5.3. Prophylactic disinfection shall be always carried out before introduction of a new group of animals to the holding or once a year;

5.4. Disinfection carpet soaked with a disinfection agent shall be placed at the entrance to the animal holding (2% Na OH solution is a universal disinfectant. In wintertime NaCl (10-15%) may be added).

#### 6. Compulsory disinfection:

6.1. The type of pathogen shall be taken into consideration upon the choice of disinfection substances, their preparation method, disinfection range and frequency as per requirements of FVS;

6.2. Disinfection shall be carried out in vehicles and premises, where the contaminated or suspected animals were kept, as well as in auxiliary premises;

6.3. All objects, mangers, equipment that have been in contact with the contaminated animal, as well as litter and manure shall be disinfected.

6.4. Final disinfection is carried out after elimination of the disease, before lifting quarantine and restrictive measures as per instructions on elimination of the given disease.

6.4.1. Animal holdings, auxiliary premises, surroundings, animal care equipment, personnel clothing, sewage, slurry and manure reservoirs shall be disinfected;

6.4.2. Laboratory disinfection control shall be carried out after disinfection. If necessary, disinfection shall be repeated.

### IV Basic principles of disinfection of animal holdings and auxiliary premises

#### 7. Basic principles of disinfection of animal holdings and auxiliary premises:

7.1. Moisturizing of premises, litter and manure with the disinfection substance;

7.2. Cleaning, washing and scrubbing of floors and walls. Cleaning and disinfection of floors and manure canals shall be carried out thoroughly, settling reservoirs shall be emptied. The quality of mechanical cleaning is enhanced by the use of water. If the dirt cannot be washed, 2% NaOH or calcinated soda (70-80<sup>0</sup>C) disinfection solution can be used. After expiration of exposition time the solution and dirt shall be washed away with a stream of hot water. After drying of premises and constructions disinfection process can be initiated;

7.3. Treatment with disinfection substance – disinfection;

7.4. Washing or other treatment (if required by the user's instruction) to eliminate the threat of possible contamination of animals with disinfection residues;

8. In case of necessity sanitary recondition shall be carried after the disinfection (renovation of damaged floor covering, feeding tables, etc.). Repeated disinfection shall be carried out after reconstruction.

## **V Rehabilitation of manure**

### **9. Types of manure rehabilitation:**

- 9.1. Chemical treatment;
- 9.2. Physical treatment.

10. *Chemical treatment of manure* – by mixing with one of disinfection substances active against the given pathogen;

### **11. Physical treatment of manure:**

- 11.1. Dry manure can be destroyed by burning;
- 11.2. Biothermal treatment is an effective method for manure rehabilitation.

### **12. Biothermal treatment of manure:**

12.1. Base for manure shall be prepared. The base shall be made of concrete (hard clay or wooden base is also acceptable), slightly slant with a cavity for liquid. Width – 2.5 to 3 m, length – depending on the amount of manure, height – up to 2 m. Air circulation at the bottom of the pile shall be secured by installing groove in the base;

12.2. The base shall be covered with a 40 cm layer of peat, sawdust or straw;

12.3. Manure shall be piled;

12.4. The pile shall be covered with at least 20 cm of straw, sawdust or peat for heat isolation;

12.5. The pile shall be covered with a water-resistant material (polythene) for protection from excess humidity. Before the biothermal treatment the humidity of manure shall be at least 70%, if it is lower, the manure shall be moistened;

12.6. Time of exposition depends on persistence of the pathogen. On average in summer it is 2 months, in winter – 3 months starting from the moment when the temperature has reached 60<sup>0</sup>C at the bottom of the pile. Temperature control shall be initiated approximately in a week.

12.7. Liquid that appears as a result of biothermal treatment shall be treated with chloride of lime to eliminate further spread of the disease;

12.8. After biothermal treatment manure can be used for field fertilisation.

## **VI Burning and burying of carcasses**

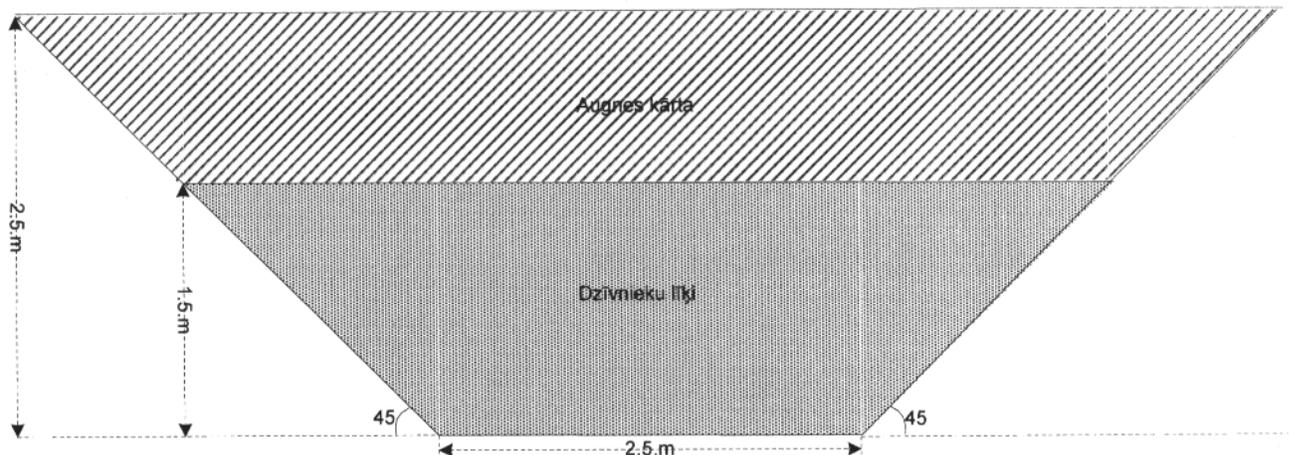
13. If the corpse is to be buried, then:

13.1. The place of burial shall be approved by the regional environment protection inspector. Location of nearby water bodies and the level of ground-water shall be taken into account.

13.2. Before burying the carcasses shall be treated with a disinfection agent;

13.3. For preparing the burying place – see picture;

### Preparation of burying place



14. If the corpse is to be burned, then burning shall be approved by the regional fire and rescue service;

14.1. **Pyre burning** is an open air system of burning carcasses either on – farm or in collectives' sites fueled by additional materials of high energy (approximately 10 liters of fuel and 2.5 m<sup>3</sup> of wood are necessary for burning of one cattle). Because this process can be conducted on site there is no requirement for transportation of the input material. The process is contrary to environmental standards for air, water and soil. The process takes the extended period of time. This is uncontrolled process with no verification of pathogen inactivation.

14.2. **Mass burial or open farm burial** is a system to deposit whole carcass below ground level and to be covered by soil (or Mounding – mass burial above ground level, with no additional inactivation of pathogens. This procedure is conducted on site, does not require transportation and is used to control the spread of disease. The process requires an environmental assessment because of the potential contamination of ground water. The environmental impact increases with the number of carcasses. The process does not inactivate all pathogenic agents. The site of burial is lost for future use for an extended period of time.

### VII Treatment or destruction of animal fodder and litter

15. Animal fodder and litter can be:

- 15.1. burned;
- 15.2. treated biothermally together with manure;
- 15.3. treated with a disinfection agent.

### VIII Disinfection of slurry and sewage

16. In case of necessity sewage and slurry shall be disinfected by adding disinfection agent, e.g. 3 kg of formaldehyde to 1 m<sup>3</sup> of the liquid or chloride of lime containing 25% active chloride, using 25 kg to 1 m<sup>3</sup> of the liquid. Time of exposition should be 72 hours. Milk not for animal or human consumption can be treated similarly.

### IX Rehabilitation of soil

17. In case of necessity as per instruction of the veterinary doctor soil disinfection measures shall be applied taking into consideration specific characteristics of the pathogen. Usually the surface is treated with a disinfection substance and then ploughed or buried.

### **X Disinfection of vehicles**

18. Upon treating the outer parts of vehicle with a disinfection substance, high-pressure disinfection equipment (hydropanels) can be used.

19. All vehicle equipment including ropes and tethers used for animal tying shall be washed and disinfected.

20. Tyres shall be disinfected thoroughly. Vehicles shall go through the disinfection barrier tyres making at least two turns. Special disinfection barriers, where substrate soaked with a disinfection substance is used (sawdust, etc.) can be set up. Hydropanels can be used.

### **XI Disinfection of personnel clothing and footwear**

21. In case of suspicions or outbreak of an infectious animal disease, experts, farmers, members of family, workers and other persons within the affected territory shall disinfect clothing and footwear.

22. Especially created disinfection cameras – formaldehyde steam cameras, as well as various chemical disinfection substances can be used for disinfection.

23. Clothing is treated by moistening with the disinfection agent and then washing carefully.

**Principles of choice of traditional chemical disinfection substances**

Disinfection substance	Pathogen groups and concentration of disinfectants (%)			
	I <sup>(1)</sup>	II <sup>(2)</sup>	III <sup>(3)</sup>	IV <sup>(4)</sup>
<b>Natrium alkali</b>	<b>2</b>	<b>2</b>	<b>3</b>	<b>10</b>
<b>Formalin, paraformaldehyde</b>	<b>2</b>	<b>2</b>	<b>3</b>	<b>4</b>
<b>Chloride of lime</b>	<b>2</b>	<b>3</b>	<b>5</b>	<b>5</b>
<b>Glutaraldehyde</b>	<b>0.5</b>	<b>1</b>	<b>1</b>	<b>2</b>
<b>Lysol</b>	<b>5</b>	-	-	-
<b>Freshly slaked lime suspension</b>	<b>20</b>	<b>20</b>	<b>20</b>	-
<b>Calcinated soda</b>	<b>5</b>	-	-	-

**<sup>(1,2,3,4)</sup> Division of pathogens in four groups:**

- <sup>(1)</sup> of low persistence: enzootic bovine leucosis, brucellosis, colibacillosis, leptospirosis, lysteriosis, Aujeszky's disease, pasteurellosis, salmonellosis, trichomoniasis, campylobacteriosis, trypanosomiasis, toxoplasmosis, infectious rhinotracheitis, parainfluenza, bovine viral diarrhoea, infectious agalactia, sheep and goat contagious pleuropneumonia, infectious atrophic rhinitis, dysentery, transmissible gastroenteritis, balantidiasis, erysipelothrix insidiosa, equine rhinopneumonitis, pulorosis, micoplasmosis, myxomatosis, calf diarrhoea, other diseases caused by pathogen microflora (proteuss, klebsiella, morganelle).
- <sup>(2)</sup> persistent: adenovirus infection, foot-and-mouth disease, turalemia, ornithosis, staphylococcosis, streptococcosis, rabies, plague, necrobacillosis, aspergillosis, candidiasis, trichophyton, microsporia, other mycoses, chlamydiosis, rickettsiosis, enterovirus infection, influenza, malignant head fever, actinomycosis, infectious catarrhal fever, infectious sheep mastitis, foot decay, swine vesicular disease, infectious anaemia, infectious encephalomyelitis, epizootic limphangitis, Malleus, duck viral hepatitis, goose viral enteritis, infectious bronchitis, laryngotracheitis, Marek's disease, Gumboro disease, infectious poultry encephalomyelitis, Newcastle disease, smallpox, infectious hepatitis of carnivores, viral enteritis, Aleut disease, pseudomyiasis, rabbit haemorrhagic disease, non-classified viruses.
- <sup>(3)</sup> very persistent: tuberculosis, paratuberculosis.
- <sup>(4)</sup> especially persistent: Anthrax, lamb dysentery, piglet anaerobe enterotoxemia, Coccidiosis, Bradzot, emphysematous carbuncle, infectious sheep enterotoxemia, other pathogens producing spores.

**Disinfection and rehabilitation measures in case of an outbreak  
of Foot-and-mouth disease**

1. Persistence of pathogen (virus). Persistence of the pathogen depends on the temperature and type of substrate, where it is found. In dry environment, darkness and cool the virus can last up to 2 years. The pathogen is sensitive to pH changes, can be destroyed at pH <6 and >10.
2. Chemical disinfection substances. All disinfection substances active against foot-and-mouth disease pathogen and registered in the Republic of Latvia, as well as traditional substances listed in the annex, can be used for disinfection. Foot-and-mouth disease virus can be easily destroyed by alkali or acid. It is important to observe user's instructions. Alkaline disinfection substances should be heated till 70-80°C.

Elimination of foot-and-mouth disease pathogen  
in products and materials of animal origin

1. Elimination of the virus in meat products:

- 1.1. In preserves:

The meat shall be treated thermally in a closed jar, so that the inner temperature is at least 70°C for 30 minutes. An equivalent method securing elimination of the virus is acceptable.

- 1.2. Thermal treatment of meat and meat products:

Meat without bones and fat shall be treated thermally, so that the inner temperature of the product is at least 70°C for 30 minutes. Then the meat shall be packed in a way to prevent repeated contamination.

- 1.3. Meat drying after salting:

When "rigor mortis" is finished, bones shall be removed, meat shall be salted and dried completely. Drying means that the water protein proportion shall not exceed 25.5:1.

2. Disinfection of raw skin. Before industrial processing skins shall be salted in sea salt, containing 2% natrium carbonate.

3. Elimination of FMD pathogen in milk and cream for human consumption (any of the below methods can be chosen):

- 3.1. Treatment at an especially high temperature of 132°C for at least one second;

- 3.2. If milk pH is lower than 7 pasteurisation shall be carried out (72°C at least 15 seconds);

- 3.3. If milk pH is higher than 7, pasteurisation shall be performed twice.

4. Elimination of FMD pathogen in milk for animal consumption (any of the following methods can be chosen):

- 4.1. Repeated pasteurisation at 72°C for at least 15 seconds;

- 4.2. Pasteurisation combined with any other physical treatment, e.g. sustain product pH level at 6 for at least 6 hours or combine with milk condensation;

- 4.3. Combining treatment at a very high temperature of 132°C with any other physical method, e.g. as per 4.2.

5. Treatment of skin and trophies of FMD susceptible animals (any of the following methods can be chosen):

- 5.1. Boiling in water till soft tissue can be removed easily;

- 5.2. Soaking in 4% Na<sub>2</sub>CO<sub>3</sub> solution securing pH level not lower than 11.5 for at least 48 hours and mixing the solution;

- 5.3. Soaking in formic acid solution (100 kg NaCl and 12 kg formic acid to 1000 litres of water), securing pH level not lower than 3 for at least 48 hours and mixing the solution;

- 5.4. Raw skin shall be salted in sea salt containing not less than 2% Na<sub>2</sub>CO<sub>3</sub>.

6. Elimination of FMD pathogen in wool for industrial processing (any of the following methods can be chosen):
  - 6.1. Industrial washing that includes repeated dipping in water basin, soap and soda or potassium hydroxide solution;
  - 6.2. Chemical depilation with slaked lime or sodium sulphide;
  - 6.3. Treatment with formaldehyde in a hermetically closed camera for 24 hours. The most practical method is inserting potassium permanganate in a container (cannot be made of plastic or polyethylene) and adding commercial formalin. Approximately 53 ml of formalin and 35 g of potassium permanganate shall be added per one cubic metre of the camera.
  - 6.4. Storage of wool at a temperature of 18<sup>0</sup>C for four weeks, at 4<sup>0</sup>C – four months or at 37<sup>0</sup>C – 8 days.
  - 6.5. Industrial cleaning, which includes dipping of wool in a hot, water-soluble detergent at a temperature of 60-70<sup>0</sup>C.

## Traditional disinfection substances and necessary concentration for elimination of FMD virus

Disinfection substance in	Carpets	Premises, holdings	Equipment containers	Slurry, sewage	Carcasses	Hands	Surroundings, soil	Manure	Clothing, footwear
Natrium alkali NaOH	2%	2% 1l/m <sup>2</sup> 24h	2% 24h	-	2%	-	2% 10l/m <sup>3</sup> 24 h	-	2%
Calicanted soda Na <sub>2</sub> CO <sub>3</sub>	5%	5% 1l/m <sup>2</sup> 24h	1-2% 24h	-	5%	-	-	-	1-2%
Parasoda	1-2%	1-2% 1l/m <sup>2</sup> 24h	1-2% 24h	-	1-2%	-	-	-	-
Chloramine-B C <sub>4</sub> H <sub>5</sub> SO <sub>2</sub> NClNa X 3H <sub>2</sub> O	-	-	2% of active Cl	-	-	2% of active Cl	-	-	2% of active Cl
Formaldehyde <sup>1</sup> HCH	-	2-4% 1 <sup>0</sup> +15 <sup>0</sup> -30 <sup>0</sup>	2-4%	2-4%	2-4%	-	2-4% 10l/m <sup>3</sup> 24 h	-	-
Potassium hydroxide KOH	2%	2% 1l/m <sup>2</sup> 24h	2%	-	2%	-	2% 10l/m <sup>3</sup> 24 h	-	-
Quick lime Ca(OH) <sub>2</sub>	-	Freshly slaked 20% 1.5l/m <sup>2</sup> 3x with 2 h interval	-	-	20%	-	5kg/m <sup>2</sup>	-	-
Chloride of lime <sup>2</sup> Ca(OCl) <sub>2</sub>	2% of active Cl	5% of active Cl 5 h	2-5% of active Cl	25%	25% of active Cl for 24h	-	25% 10l/m <sup>3</sup> 24 h	25% 24 h	-
Hypochloride <sup>3</sup>	0.5% of active Cl	10% of active Cl 0.5l/m <sup>2</sup> 3h	10% of active Cl	10%	-	From 0.5% of active Cl	-	-	-
Sulphonamide acid <sup>5</sup>	-	-	-	-	-	0.2%	-	-	-
Citric acid <sup>4</sup>	-	-	-	-	-	0.4%	-	-	-
Technical orthophosphor acid	-	-	-	-	-	0.3%	-	-	-

**Formaldehyde HCH**

Colourless gas, which dissolves in water. Upon disinfection with formaldehyde gas the temperature in premises should be 15-30°C.

**<sup>2</sup> Chloride of lime Ca(OCl)<sub>2</sub>**

The substance is available with 38, 32, 28, and 25% content of active chloride. The quality of the chloride of lime is determined based on the content of active chloride. For disinfection purposes cleared chloride of lime solution with 2-5% content of active chloride is prepared. Chloride of lime with 25% content of active chloride is usually mixed with slurry and sewage.

**<sup>3</sup> Hypochloride** – its activity decreases during storage, therefore it is necessary to check it before the use. It is a disinfection substance with a wide spectrum of application.

**<sup>4</sup> Solution of citric acid** – cannot be stored longer than for 2 weeks. Container should be closed hermetically to eliminate reproduction of acid resistant bacteria.

**<sup>5</sup> Sulphonamide acid solution** – 500 grams of crystals to 240 liters of milk is used for virus elimination in milk reservoirs and containers. As this acid has less impact on metals than other mineral acids, in 2% concentration it can be used for disinfection of metallic and painted surfaces, rubber and plastics. After disinfection the equipment should be washed carefully with water.

**DISINFECTANTS USED FOR FOOT AND MOUTH DISEASE**

Virkon S	1% (1:100); 2% (1:200)	Organic acids and catalyst
Virusan S		
Parvocide Plus	1:80	Glutaraldehyde
Citric acid	1:500	Citric acid
Capo Tabs	0,5%	Na-Cl-isocyanurate
Chloride Forte	3%	Na-hypochloride
P3-tapax 66	3%	Na-hypochloride
Jobac P	3%	Iodine
Hot caustic lye	2%, 3%	Na hydroxide
Septabic	0,1%	Urea, DDAB
Bromosept	0,1%	DDAB, ethanol
Diklo	0,5%	Na-dikloroisotsüanoraat
Desomix	1%	Na compounds, nonionogenic tensit
Deso	3%	Na compounds
Foam-Deso Vahu-Deso	4%	Na compounds, tensit
Sodium hypochlorite	1-2%	Na hypochlorite
Virocid	0,5%	

Other disinfectants may be used according to the preference described on the handout and its instruction. The listed disinfectants are also used according to the accompanying instruction.

Disinfection pools (disinfection mats)	Virkon S, Citric acid, Septabic, Bromosept Diklo, Sodium hypochlorite, Virocid
Initial disinfection	Virkon S, Virusan S
Impact time	24 hours
Washing	Pintty-detergent (Farmos)
Minimum break	7 days
Final disinfection	Virkon S, Parvocide Plus, Virusan S
Impact time	24 hours

Minimum break	7 days
Disinfection of the personnel	Citric acid, Virkon S sidrunhape, Virkon S
Clothing	Virkon S. Citric acid or heat treatment (at least 70°C / 2 hours)
Animal fur	Virkon S, Citric acid
Equipment, vehicles	Virkon S, Deso, Fomay Deso, Diklo
Drinking water distribution system	Virkon S, Capo Tabs
Milk pipes and cooling tanks	Virkon S, Capo Tabs, Diklo, Desomix
Milk	Will be poured into the liquid manure storage facility or pit
Solid manure	Lots of unslaked lime will be sprayed on its surface
Liquid manure	At least 30 kg of unslaked lime per one ton of manure will be added, permanently mixing for at least 6 hours
Liquid manure pits	At least 30 kg of unslaked lime per 1 m <sup>3</sup>
Storage of concentrated feed	Virkon S, caustic lime
Feed (hay, straw, flour et al.)	Virkon S
Air in the livestock buildings	Virkon S
Ground	Virkon S, caustic lime

## **Disinfection and Decontamination Measures in Case of Classical Swine Fever Outbreak**

**1. Agent (virus) resistance.** The CSF resistance depends upon the ambient temperature and the medium where the disease agent is found. The virus is resistant in the outside environment at low temperatures (in +4°C 4-6 months). In frozen pig meat it may remain active for several years; in salted pig meat – up to 4 months. The virus is rapidly inactivated in sunshine, elevated temperatures (+70°C) and appropriate disinfection products.

**2. Chemical Disinfection Products.** Any disinfection product registered in the Republic of Latvia and efficient in respect of agent of Classical Swine Fever or any of the traditional products listed in the table to this Annex may be used for disinfection. The disinfection should be carried out in accordance with instructions of the veterinary inspector. The activity of disinfectants is to be checked before use, as activity of certain disinfectants is diminished by prolonged storage. The directions for use should be closely followed.

**3. General Principles of Disinfection**

**The following activities should be carried out:**

- 3.1. Thorough soaking of bedding and litter as well as faecal matter with the disinfectant;**
- 3.2. washing and cleaning by careful brushing and scrubbing of the ground, floors, ramps, walls and other objects; the water used for cleaning operations is to be disposed of in such a way as to avoid any risk of spreading the virus;**
- 3.3. This should be followed by application of disinfectant to above objects, fittings, installations and other possibly contaminated objects. Where disinfection equipment is used with liquids applied under pressure, re-contamination of the previously cleansed parts is to be avoided**
- 3.4. Following the disinfection procedures, re-contamination is to be avoided;**
- 3.5. The performed cleansing and disinfection measures have to be documented.**

**4. Special provisions on cleansing and disinfection of infected holdings.**

**4.1. Preliminary Cleaning and Disinfection:**

- 1) During the killing of the animals all necessary measures are to be taken to avoid or minimize the dispersion of classical swine fever virus. This is to include inter alia the installation of temporary disinfection equipment, supply of protective clothing, showers, decontamination of used equipment, instruments and facilities and the interruption of power supply to the ventilation;**
- 2) carcasses of killed animals are to be sprayed with disinfectant;**
- 3) removing carcasses to the place of disposal, covered and leak proof containers are to be used to avoid the contamination of environment;**
- 4) as soon as the carcasses of the pigs have been removed for processing, the holding and the possibly contaminated environment should be immediately sprayed with disinfectant;**
- 5) any tissue or blood which may have been spilled during slaughter or post-mortem or gross contamination of buildings, yards, utensils, etc., is to be carefully collected and processed with the carcasses;**
- 6) The disinfectant used is to remain on the treated surface for at least 24 hours.**

**4.2. Final cleansing and disinfection:**

- 1) manure and bedding are to be removed, stacking and dampening (if necessary to**

facilitate fermentation), covering to keep in the heat so that a temperature of 20 °C is attained and leaving covered for 42 days; they may be disposed of also by burying or incineration;

- 2) grease and dirt are to be removed from floors and other surfaces by the application of a degreasing agent and the surfaces washed with water;
- 3) disinfectant must be applied to all surfaces;
- 4) after seven days the premises are to be treated with a degreasing agent, rinsed with water, sprayed with disinfectant and rinsed again with water;
- 5) slurry is to be stored for at least 42 days after the last addition of infective material, unless the competent authorities authorize a reduced storage period for slurry which was actually treated in accordance with the instructions given by the official veterinarian so as to ensure the destruction of the virus.



## Disinfection and Decontamination Measures in Case of Newcastle Disease Outbreak

**1. Agent (virus) resistance.** The agent is inactivated at 56°C in 1.5 hours. At temperature -20°C, the virus is active for the period of up to 20 years. In pH environment from 2 to 10, the agent retains virulence up to 10 days.

**2. General Cleaning and Disinfection.** As soon as the carcasses of the poultry have been removed for disposal, those parts of the premises in which the poultry was housed and any parts of premises, where activities with birds have been carried out, as the result of which the Newcastle disease agent may have been spread, should be sprayed with a disinfectant registered in Republic of Latvia, effective in respect of Newcastle disease agent.

Any tissue of poultry or other contaminated material which could have contaminated buildings and yards should be carefully collected and disposed of with the carcasses. The used disinfectant must remain on the surface for at least 24 hours.

### 3. Final Cleaning and Disinfection.

Grease and dirt should be removed from all surfaces by the application of a degreasing agent and washed with water.

After washing with water, further spraying with disinfectant should be applied.

After seven days the premises should be treated with a degreasing agent, rinsed with cold water, sprayed with disinfectant and rinsed again with water.

Used litter and manure must be treated by a method capable of killing the virus. This method must comprise at least one of the following practices:

- incineration or steam treatment at a temperature of 70 °C
- burying deep enough to prevent access by vermin and wild birds;
- stacking and dampening (if necessary to facilitate fermentation), covering to keep in the heat so that a temperature of 20 °C is attained and leaving covered for 42 days so as to prevent access by vermin and wild birds.

NB! Newcastle disease agent is susceptible to **traditional** disinfection products, listed in the table of Annex 3 to this instruction.

## **DISINFECTION AND DECONTAMINATION MEASURES IN CASE OF AVIAN INFLUENZA OUTBREAK**

### **1. Preliminary Cleaning and Disinfection:**

1.1. As soon as the carcasses of the poultry have been removed for disposal, those parts of the premises in which the poultry was housed and any parts of other buildings, yards etc. contaminated during slaughter or post-mortem examination should be sprayed with a disinfectant registered in the Republic of Latvia.

1.2. Any tissue of poultry or eggs which could have contaminated buildings, yards, utensils etc. should be carefully collected and disposed of with the carcasses.

1.3. The used disinfectant must remain on the surface for at least 24 hours.

### **2. Final Cleaning and Disinfection:**

2.1. Grease and dirt should be removed from all surfaces by the application of a degreasing agent and washed with water,

2.2. Subsequent to removal of grease and dirt prescribed under point 2.1., further spraying with disinfectant should be applied;

2.3. After seven days the premises should be treated with a degreasing agent, rinsed with cold water, sprayed with disinfectant and rinsed again with water.

2.4. Used litter and manure must be treated by a method capable of killing the virus. This method must comprise one of the following practices:

2.4.1. incineration or steam treatment at a temperature of 70° C;

2.4.2. burying deep enough to prevent access by vermin and wild birds;

2.4.3. stacking and dampening (if necessary to facilitate fermentation), covering to keep in the heat so that a temperature of 20° C is attained and leaving covered for 42 days so as to prevent access by vermin and wild birds.